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PATHOGENS OF SELECTED MEMBERS
OF THE PAPAVERACEAE--
AN ANNOTATED BIBLIOGRAPHY

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ACKNOWLEDGMENTS

The authors hereby respectfully acknowledge the encouragement and helpful assistance of Quentin Jones, Donald Fishler, Paul Lentz and staff, Charles Kingsolver, Kenneth Bromfield, Charles Beebe, and members of the National Agricultural Library staff; Sue Gorman and Robert Freytag for a number of translations; and Hilde McGrath and staff for considerable assistance on the abstracts.

PATHOGENS OF SELECTED MEMBERS OF THE PAPAVERACEAE--AN ANNOTATED
BIBLIOGRAPHY

By Chris G. Schmitt and Bernard Lipscomb^{1/}

INTRODUCTION

This annotated bibliography presents a brief review of some of the more important contributions on pathogens of several members of the Papaveraceae. Although the bibliography is quite extensive, we realize that it is not complete. A work of this type can never be completely finished because of continuous efforts on the part of researchers in all parts of the world. Our sincere apologies are offered to any whose contributions have not been included. In some instances omissions are due to lack of availability and in others to a language barrier.

Because of frequent occurrence of the genus name *Papaver* in the review and bibliography, we have abbreviated that genus name with a *P.* to conserve time and effort.

National Agricultural Library call numbers are included in the bibliography citations to assist those who desire to obtain original articles.

^{1/} Respectively, plant pathologist, Plant Disease Research Laboratory, Northeastern Region, and agricultural research technician, Mycology Laboratory, Plant Protection Institute, Agricultural Research Service. Mr. Lipscomb is chiefly responsible for the bibliography, an integral part of this report.

DISEASES CAUSED BY FUNGUS PARASITES OF PAPAVERACEAE

Parasitic Leaf Drying, or Poppy Fire

Taxonomy

Pleospora papaveracea (de N.) Wint., 1887 (604) 2/

Syn.: *Cucurbitaria papaveracea* de N., 1863 (468).

Pleospora papaveracea (de N.) Sacc., 1883 (468).

Sphaeria pellita Fr., 1821.

Pyrenophora pellita (Fr.) Sacc., 1883 (325, 468).

Pleospora pellita (Fr.) Rab., 1887 (325, 368).

Sphaeria calvescens Fr., 1843.

Pleospora calvescens (Fr.) Tul., 1863.

Pyrenophora calvescens (Fr.) Sacc., 1883.

Imperfect stage, chiefly responsible for parasitism.

Dendryphon ('*Dendryphium*') *penicillatum* (Cda.) Fr., 1840
(96, 97, 121, 617).

Brachycladium penicillatum Cda., 1838.

Helminthosporium papaveris P. Henn., 1907.

Helminthosporium papaveris (as '*papaveri*') Saw., 1918
(325, 498).

Dendryphium papaveris (Saw.) Saw., 1959.

Dendryphium ramosum Cooke, 1871 (498).

Distribution and Hosts

Especially common in Europe on *P. somniferum*, poppy fire has been found on this host in the Orient and in Australia. It has been seen upon *Chelidonium majus* L. and there are scattered reports of its occurrence upon several *Papaver* species. As a pathogen, the fungus is closely restricted to species of *Papaver*, but as a saprophyte it can grow on various genera (23).

Symptoms

Seedlings attacked may die before emergence when inoculum is present on the seed. On diseased plants that emerge there is root rot, browning, and constriction of the xylem. Roots and even whole plants may turn black. Before and during blossoming, mass infection of leaves develops resulting in numerous black-brown to black angular spots. Blue-black striping appears on stems, leaf bases, and veins. Lower leaves wither. With high humidity

2/ Underscored numbers in parentheses refer to bibliography, p. 31.

sporulation occurs and spread is rapid. Flower heads can also wither and become black-brown. The discoloration spreads to the capsules from diseased peduncles. If capsules are infected early, they are deformed, remain small, become brown, and there is no yield. Seed on carpel walls may be covered by mycelium and conidia. Seed adhere tightly to capsule walls. Tracheo-mycosis was a feature of all diseased plants examined. Infections during the rosette stage gradually constrict the crown. Water and food supply are reduced; plants turn yellow and finally collapse (325, 354, 379, 395, 451).

Description

The considerable variation observed in spore sizes in response to differences in environment and substrate are indicated by measurements of various investigators and helps to explain the multiple epithets applied to this fungus. Measurements of spores and fruiting structures by various investigators are given in table 1.

After an intensive study upon conidial size and septation, Ballarin (23) summed up the cause of variation in his statement that conidia from poorly nourished cultures were small and had few septae compared with the large multiseptate conidia from well-nourished cultures or from poppy capsules. Schmiedeknecht (509) made similar observations. Khristov (235) and Ballarin (23) demonstrated protoplasmic connections between individual cells of conidia at the center of septae. Chlamydospores are terminal and intercalated, single and in chains. On carrot agar they averaged $12.6 \times 12.9 \mu$ (325).

That the imperfect stage can be derived from the germination of ascospores has been demonstrated by several researchers (150, 152, 325, 617). Based upon observations that perithecia of both *Pleospora pellita* (Fr.) Rabh. and *Pl. papaveracea* (de N.) Sacc. are glabrous, Petrak concluded that they were identical (408). Meffert found that *Pl. calvescens* perithecia were hairy and are found only on *Chenopodiaceae*. She concluded that the poppy fungus was *Pl. papaveracea* (325). Commonly conidia are 4-, 5- or 6-celled (451). According to Sawada *Helminthosporium papaveris*, *Dendryphium penicillatum* (Corda) Fr. and *D. ramosum* Cooke all occur on *P. somniferum* (504). There is no advantage in belaboring his descriptions of these since they are all the same fungus.

Conidia are produced in abundance on poppy decoction agar plus 2 percent glucose (24, 325). Physiologically acid salts (pH 5.6) inhibited and alkaline salts (pH 7.5-8.0) promoted growth in nutrient solutions. Best sources of carbon were lactose and glucose, and of nitrogen, potassium nitrate, asparagine and glycocol (325). Ballarin found nitrates and ammonium compounds were suitable nitrogen sources (23). Nature of the medium greatly influenced manifestation of characteristics of individual strains (509). Mycelial growth rate was distinctly retarded when relative humidity dropped below 98 percent (510). In liquid medium optimum pH for

TABLE 1.--*Measurements of spores and fruiting structures of Pleospora papaveracea (Dendryphion penicillatum)*

Investigator	Conidia	Conidio- phores	Perithecia	Asci	Ascospores
Ballarin (23)	5-8x30-35 μ (on malt agar) 5-8x35-70 (on capsules)	30-50 μ	300 μ (host) 70 μ (media)	90-110 μ	8x17-22 μ
Barcacka (24)	6.6-10.6x18-112		170-310 wide	10-15x	6.5-11x
Darpoux (94)	4-11x15-60	4-7x20-60	120-240 high	84-135	16.5-27
Ducos (112)			300 max. dia.	12x100	7x20
Gizhitskaia (149)	3-5x25-35			10-11x 110-150	8x22
Khristov (235)	5-8x15-57 (oat agar)				
Meffert (325)	3-7x10-22 uniseptate 7-8x31-40 quinquiseptate		140-390x 180-360	11-16x 106-134	5-10x 15.5-24
Mraz (354)	5.2-11.7x17-42			90-110	8x17-22
Sawada (498)	7-11x22-112				
Tanaka (550)	7-11x22-112	6-7x86-130			
Winter (604)				10x100	7-8x18-25

mycelial growth was near 5, minimum 3 and maximum above 10 (23). Starch and pectin were degraded (325). Cellulose was not utilized, but all sugars tried and poppy seed oil were used (23).

Epiphytology

Initiation of infection in the spring is often from seed from diseased plants (235, 325, 449 to 451) or can occur from conidia or ascospores from debris of the previous crop or from wild species of poppy (325).

In the Ukraine poppy fire appears in June and mass infection follows in July, usually at the bud stage. High humidity in June and July favor infection. No resistant varieties were found (38). Disease appeared at Seine-et-Oise in July 1944 and by August nearly all capsules had been attacked, but only the late ones were severely damaged. Infection was obtained in a saturated atmosphere and spots appeared in 3 days. Intensity of attack varied with variety (94). *P. somniferum* is more prone to attack than are wild poppy species. Strains of the fungus varied in virulence (325). Conidia are spread by wind, insects, and man; infection is favored by high temperature and humidity. Incubation period is 4 days (128). The fungus can penetrate from any point on the capsule wall (144, 324, 325). One investigator maintained that the fungus may be impeded by reasonably narrow spacing to insure rapid and uniform crop growth (144).

Ballarin (23) noted that infections were more common on border plants than on plants in the center of the stand and attributed the higher incidence upon border plants to their greater succulence and slower maturation. Plants are most susceptible as seedlings and at flowering. Infection begins on lower leaves and spreads upwards. Strongly branched plants in thin stands mature more slowly and are more severely infected than poorly branched plants in thick stands (167).

Grummer (168) observed that infection of older plants does not occur until some part of the leaves are predisposed to the fungus through protein decomposition. Weather must also be favorable for attack when this condition is met. Chlamydospores may be important in spanning unfavorable growth periods. High moisture and temperature (24 to 30° C. opt.) are necessary for an epiphytotic (128, 235, 395). Plants may be attacked in all stages of development (19, 235). Incubation period is 2 to 8 days depending upon temperature. Variability in pathogenicity of strains occurs.

Schmiedeknecht (509) demonstrated the existence of strain differences in cultures derived from many single-spore isolations.

Mraz studied health effect of heterosis in an area where the disease is a problem. He found the Czech varieties more resistant to the fungus than foreign varieties (350, 352, 353). The fungus is seed-borne on *P. somniferum* and *P. rhoeas* (374). The poppy capsule curculio, *Ceuthorrhynchus macula-alba* Herbst., breaks capsule walls either during egg deposition or larval exit.

Fungi enter and cause molding of the capsule. For example, in the summer of 1951 a field free of the capsule curculio had 7.5 percent moldy capsules but 98 percent of capsules were moldy in a field where the curculio was present (380).

Varieties in Romania vary in resistance to the disease (404). In Romania poppy fire increases in direct ratio to sowing density. In 1959 60 percent of Tatarstan 1 and 11 percent of Margurele plants were attacked (435).

Poppy fire is more prevalent in light soils and almost absent on loamy soils in Germany. In thin stands the disease was absent (450).

In north Germany incidence of the disease was markedly higher in the summer of 1941 than in 1942 because of abnormally high temperature in 1941. The disease was severe both in Sweden and in Rostock in 1941 because of the higher than normal temperatures in June and July. A high May temperature in 1942 caused an early and alarming buildup, but cool June and July temperatures dissipated the epiphytotic (450).

Incidence of poppy fire is high in the warmer countries, especially in Bulgaria where whole fields may be destroyed (395). The fungus was favored by high temperature and heavy rainfall in Czechoslovakia, especially in July (348). Not only is germination affected, but plant stand can be greatly diminished. Early establishment of conidial formation permits large spore populations for secondary infection and weather prevailing at this time exerts a deciding role upon disease development. Temperatures during June and July (25 to 30° C opt.) are quite important for disease development. Little, if any, damage occurs on seedlings in cool weather (325). Inoculations of unwounded leaves and stems of healthy poppies with conidia gave negative results even in a saturated atmosphere, but when surfaces were scratched lightly before inoculation, infection ensued in 4 to 5 days (610).

All plant parts at all stages of development are attacked by all spore forms. At lower or higher temperatures where the fungus grows slowly, the host forms demarcation zones that localize the pathogen but do not eliminate it. These gumming demarcation zones of 10 to 30 cell layers form within a week at temperatures below 16° and above 30° C. At temperatures that are optimal for the fungus the host may be overcome in several days. Resumption of favorable temperatures after a cool period permits further depredations. Conidia can germinate within 2 hours of maturity. The optimum temperature for germination is 10° to 20° C, minimum 3° and maximum 33° C. Optimum temperature for infection is 24° to 28° C, minimum 5 to 10 and maximum 34° (616, 617, 618). The earlier the poppies are sown in Romania the more reduced is the attack and the greater the production (435).

In Sweden field temperatures were higher than normal in June 1941 through early August 1941 and infection was severe. The following year temperatures were below normal for these months and poppies were not diseased although plants from the same seed lots raised in the greenhouse at higher temperatures were heavily infected (31).

Isolates of the fungus have remained viable on malt agar at 20° C for 55 months. Rate of germination decreased with age of culture--spores from recent transfers (5 days) germinated best (93). Some antibiotics produced by Actinomycetes and bacteria inhibited the fungus (96, 97, 110, 124, 559). Among its metabolites the fungus produces a widely unspecific phytotoxic substance that is inactivated by oxidation (163, 165). Grümmer (168) contends that infection appears with the initiation of protein degradation in the leaf when weather conditions are propitious so that outbreak of disease is dependent upon both internal and external factors. Susceptibility of leaves was determined by a chlorosis with aging that was associated with chlorophyll disintegration and protein degradation. Attacks of the fungus were more severe on branched indeterminate plants because the slow ripening plants were susceptible over a long period.

Economic Importance

The imperfect stage of *Pl. papaveracea* is the cause of a serious disease variously referred to as parasitic leaf drying, poppy fire, helminthosporiosis, and helminthosporiose of poppy. Ravages of the parasite are closely tied to field temperatures, relative humidity, condition of plants and varieties.

Seed production loss in Germany from an attack of average severity in 1947 was 40 percent (23). In certain fields in France in 1944 up to 50 percent of the plants were affected (94). Poppy fire is the most destructive disease of the crop in Bulgaria. Whole fields have been ruined (235). According to locality in Czechoslovakia, incidence of disease varied from 2.5 to 98.8 percent. Yields in attacked fields ranged from 655 to 997 kg/ha instead of the expected 1,000 (348, 351). The fungus is indigenous to southeast and eastern European areas where it is the greatest danger to poppy cultivation (379).

Seed yields in relation to seeding date were recorded in Romania in 1960. Seeding dates were March 21, April 6, April 19 and May 12; corresponding yields were 525, 275, 225, and 0 kg/ha. Not only were yields reduced by delayed seeding, but a concomitant reduction in seed quality attributable to degree of infection occurred (435). Yields of two resistant varieties in Romania were twice that of the two susceptible varieties over a 5-year period (434).

In Oudenbosch district of the Netherlands only 15 percent of the plants remained free from infection in 1932 (420). The pathogen was the cause of considerable decrease in percentage germination of poppy seed and of seedling injury. It was by far the most common cause of seedling damage in an assay of fungus flora on 400 opium poppy seed lots from various regions of Poland (612).

Control

Early seeding is advised, March or early April in Germany to obtain maturity before the onset of high temperatures of July and August (325). In dense stands plants mature more rapidly (don't remain succulent as long) and are less liable to attack. Plants around borders were more severely attacked. Increases in seed yield were found with increases in density from 12 to 22 plants per square meter. In densely planted poppies 63.7 percent of capsules harvested were healthy, but only 11.1 percent were healthy in the outer rows (23). Early seeding and dense stands reduce seed losses (23, 435). Resistant varieties are recommended (435). Formalin at 0.25 percent as a seed treatment was effective. Bordeaux was not regarded as economical as a field spray (24). Seed treatment with Uspulun and Germisan, Gramisan, TMTD, or Agrosan was effective (30, 31, 435). TMTD and Germisan appeared to stimulate seed germination (435). Darpoux (95) advised sanitation, including burning of plant debris after harvest; practice of 3 to 4 years' rotations; treatment of seed with 300 to 400 g/hectoliter of organic mercurials or copper carbonate. Similar control measures were advocated by Flachs (128).

Grummer (164) suggested cultivation of poppies along with cover crops of carrots, beets, and yellow clover to limit the disease. This effects the uniform growth and maturity Gassner advocated through seeding poppies and carrots adjacent to a dense poppy stand (144). Antibiotic containing filtrates of Streptomyces were effective in controlling helminthosporiosis through seed treatment. Filtrates of *Penicillium patulum* and of *Penicillium expansum* were effective but were phytotoxic when the active ingredient (patulin) at 200 ppm was too concentrated (251). Direct attacks of streptomycete strains upon *D. penicillatum* hyphae have been observed (364).

Either Captan or Thiram at 0.5 percent was effective as a seed treatment in the Netherlands (289). The main infection of seed poppies occurs in the Netherlands after the crop is cut and stands drying in the field. During wet weather 50 percent of the seed may be contaminated in this manner without loss of yield (290). Capsules should be dried rapidly after harvest to prevent deterioration of seed by fungus overgrowth that often occurs with slow drying (23). Losses were reduced with applications of lime (CaCO_3 10 dz/ha) but KCl in the absence of lime on loess soil at Jena resulted in heavy infection (313).

Disease severity was reduced in the Ukraine by soil amendments of borate and manganese superphosphate (30 kg/ha) (336). Agronal at 0.5 kg/100 kg seed was an effective seed treatment and was not phytotoxic to seedlings even at twice this dosage (349). Thiram 80 percent at 200 to 400 g/100 kg seed was effective (534). Copper oxyquinolate (18 percent Cu at 600 g/hl of seed prevented spore germination and was not phytotoxic on treated seed stored for a year after treatment. Ferbam at 700 g/hl gave good control but was less effective than the copper compound. The best liquid treatment was 0.02 percent unalcylthydroxymercury and was least phytotoxic to seed of the organic mercurials tried (610). Flachs (128) recommended a protective spray

with copper-containing fungicides on seedlings. Flanderikova observed phytotoxicity to poppy with all organic tin fungicides tried, with Cuprosan Super D, Morestan, the decafentins, and Dithane M-45. The best fungitoxic effect was obtained from Polyram Combi and Polyram Ultra (132). The antibiotic, Fytostrep was not phytotoxic but Fytostrep plus CuSO_4 was. Fungicidin plus actidione at 0.05 g/l improved the health condition of poppies, but dosages in excess of this reduced yield (131).

Downy Mildew or False Mildew

Taxonomy

Peronospora arborescens (Berk.) de By., 1863 (34, 99).

Syn.: *Botrytis arborescens* Berk., 1846.

Botrytis grisea Ung., 1847 (220).

Peronospora arborescens (Berk.) de By. f. *papaveris*
somniferi Gaum., 1923 (145, 220).

Peronospora effusa Rab. var. *papaveris* Desm., (145, 220).

Peronospora effusa Rab. f. *papaveris* Fckl., 1863 (34, 145).

Peronospora grisea b *minor* Casp., in Rab., 1856 (34, 99).

Peronospora papaveris Tul., 1856 (34, 99, 220, 471).

Distribution

Peronospora arborescens is nearly world-wide in distribution and occurs on most species of *Papaver* and on species of *Argemone*, *Chelidonium*, and *Meconopsis*.

Symptoms

Heading plants exhibit bleaching in definite areas of the upper surfaces of the leaves, tumors, in-curling of leaf margins, intumescences, and bending of the stem, as well as deformation of flower buds and capsules. A gray-violet furry growth develops on the under surface of leaves and infected parts of the stem after exposure to dew or rain. Irregular, brown angular spots, usually bounded by the veins occur primarily on the lower leaves of older plants. These spots can cause an early drying of leaves; in this case mold formation seldom occurs (379, 395). Characteristically lower leaves are infected first and infection gradually moves to the upper leaves. Top leaves may remain quite healthy for sometime after the lower leaves are dry (219).

Description

Conidiophores emerge in groups of 5 to 6 from stomata on abaxial leaf surfaces and are 7 to 10 times dichotomously branched. Terminal branches are attached at an angle of 90° or more. Oospores are enveloped in a brown, weakly plicated exospore (33, 354). Aerial tissues of diseased plants may contain oospores and they may occur in a homothallic mycelium (26). Measurements of spores and fruiting structures made by various investigators are presented in table 2.

Epiphytology

Fungus mycelium in the endosperm can initiate infection of seedlings and can account for initiation of infection in the spring in some instances. Cotton (89) holds that the fungus was brought to Britain in infected seed material. Even if not present in seed itself, fragments of infected capsules in the seed lot can harbor the fungus (94, 220, 394). Oospores in capsules and seeds transmit the disease (417). Physiological races are postulated (26, 27, 145, 395, 524). Definite differences in susceptibility were observed in the varieties of oil poppy grown in the Danube lowlands of lower Austria in 1927 (395). The minimum temperature for germination of conidia is 4° to 7° C, optimum 17° to 18° and maximum 25° to 26° C. Incubation period in April and May at 12° to 14° C was 4 to 5 days (78, 220). Germination occurred on leaves at 44 to 56 percent relative humidity, but not on glass slides at this relative humidity. It occurred on glass slides only when spores were in contact with water (220).

Downy mildew appears in India about mid-February when the crop is maturing (78).

In Serbia in 1927 attacked plants were dry before time for incision. Attack began in early May and a month later the lower leaves were completely dry (219). Downy mildew often appears before winter in fall-sown plants. Those infected in the cotyledonary stage are usually killed. New infections occur during the whole period of growth, all aerial parts are equally susceptible. When the growing point is infected, the disease becomes systemic and subsequently formed leaves are infected (220). It has been shown that ultraviolet light in nature kills conidia in about 2 hours on a clear day and is regarded as a factor in epiphytology (106). Leaf spots appear in 6 to 9 weeks of sowing. Conidia germinate in 2 hours in free water. Attempts made by Behr (26) to germinate oospores were futile so he concluded that oospores play no role in epiphytology. That this was an unwarranted conclusion can be confirmed by many who have tried to induce oospore germination in this and related fungi. Kothari and Prasad (259) contend that oospores provide the main source for recurrence of the disease. If they are buried at an inch depth in soil before sowing they cause infection in seedlings. They are infective in the season following their formation and do not remain viable beyond 3 years.

TABLE 2.--*Measurements of spores and fruiting structures of Peronospora arborescens made by various investigators*

Investigator	Conidia	Conidiophores	Oogonia	Oospores
Berlese (34)	20-24x16-20 μ	10-12x300-850 μ	-	30-34 μ
Butler (71)	20-25x18-22 μ	10-12xca 1000 μ	32	21-32
Chattopadhyay (78)	-	-	33	22-23
Darpoux (94)	23.2x20.8 extremes 25x23 and 18x17	500 μ	-	-
DeBary (99)	13-13.1	-	-	-
Flachs (128)	16 μ	700-850 μ	-	-
Josifovic (220) (Yossifovitch)	oval 13-26x11-19.5 av. 18.7x16.3 elliptical 16-29x14-21 av. 23.3x16.3	-	- 30.5-53 mostly 37-41	- 23-39 24-26
Lindquist (282)	13-24x12-22	550-600x14-16	-	-
Losa Espana (287)	22x19	240	-	-
Montoro Guarch (346)	22-24x16-20	-	-	25-19.6
Mraz, et al. (354)	16	850	-	23-29 mostly 24-26
Noumov (Naomoff) (366)	12-30x13-17	10-12x300-850	40-60	30-38

Wild *Papaver* species are a factor in epiphytology (418). Studies in Poland in 1953, 1954, and 1955 revealed that infections were highest in the spring and fall and were favored by the high humidities and low temperatures of these months (417). Concentrations of alanine, leucine, phenylalanine, norleucine, glutamic acid, proline, methionine, tryptophane and tyrosine were lower in diseased than in healthy leaves (513).

In general, greatest damage is through killing of seedlings and sometimes of older plants. Reduction of yields of seed per capsule occur with a concomitant reduction of germination capacity (417). Damage can be appreciable but is dependent upon environmental conditions and host varieties. The most serious disease of poppy in India is downy mildew (20). Cool, rainy weather at seed germination results in extensive damage by *P. arborescens* in Germany (26, 27). In years of serious infestation opium production has been reduced by 50 to 60 percent (46). State Farms Liscwo near Malbroc, in Poland experienced a 60 to 70 percent destruction of plants at beginning of blooming over a 5 hectare area in 1953 (417). In Egypt *P. arborescens* attacks when plants are almost fully grown and lower leaves are damaged most (57). In 1871 losses in yield in India amounted to 30 to 40 percent.

Mildew occurs in a mild form throughout the chief opium-growing tract in Bihar and the east of the United Provinces every year. In many years localized attacks of greater severity, and occasionally epidemic outbreaks, have affected considerable areas. In the moist climate of lower Bengal, the ornamental varieties of *P. somniferum* suffer so much that they are not grown in the Calcutta Botanic Garden. In this area *P. rhoeas* is quite resistant (71). From one-third to two-thirds of the entire crop (and occasionally the entire crop) can be destroyed by downy mildew in Serbia and Yugoslavia (219, 220, 394).

In 1949 downy mildew caused severe damage in Austria because of high rainfall and frequent dew (508). Heavy losses of opium occurred on heavy, swampy land in Iran in 1970, but there were only traces of disease on well-drained soil (505). Downy mildew reduced opium yields 5 to 20 percent and seed yields by 50 to 70 percent in Russia in 1961 (621). There was little damage from it in Germany in 1898 (243). In India it accounts for considerable loss in seed and opium yield (257). Downy mildew is regarded as a serious pest in Austria (427), the Netherlands (418), Poland (417), and India (103).

Control

Dithane Z-78 at 0.4 percent applied as a drench to the soil before sowing checked infection by oospores (256). Using clean seed from healthy plants, roguing of infected plants, burning debris of the infected crop, covering debris by deep ploughing, and rotating poppy every 3 years have been recommended (89, 94, 220, 239, 259, 394, 395, 417, 427). Destroy *Argemone*

mexicana L. in fields near cultivated poppies and spray with 0.3 percent Blitox-50 or with Dithane Z-78 (308). Timely spraying with "Bayer"-mixture, 1 percent copper oxychloride, was recommended in Bavaria (128). Avoid moist areas and dense stands (379, 394, 395, 417). Resistant varieties occur and can be used to reduce losses (103), e.g., infection varied from 15 percent on Burgenland to 60 to 70 percent on Esterhaza in Austria in 1927 (427).

Gray-Mold Blight

Taxonomy

Botryotinia fuckeliana (de By.) Whetzel, 1945 (354).

Peziza fuckeliana de By., 1866.

Sclerotinia fuckeliana (de By.) Fckl., 1869.

Imperfect state: *Botrytis cinerea* Pers. ex Fr., 1821 (121).

Distribution and Hosts

Although the fungus is cosmopolitan, reports of gray-mold blight upon members of the Papaveraceae are quite rare. It has been noted on these hosts in Eurasia, United States, and New Zealand.

Symptoms

In cool, humid weather gray mold may appear on leaves, stems, flowers, and fruits. Buds and capsules may abort. It affects all growth stages. Seedlings may damp-off. Spots on stems, capsules, and leaves become dry. Infection often begins at a leaf axil and proceeds up the peduncle. Sepals become black, capsules may be deformed (354).

Description

Colonies gray or grayish brown. Production, sizes, and shapes of sclerotia on natural substrata and in culture are extremely variable. In culture some strains form no sclerotia; in others they are abundant. Sclerotia are black, usually smaller and thinner than those of *Whetzelinia sclerotiorum* Korf and Dumont. Conidiophores frequently 2 mm or more long, mostly 16 to 30 μ thick, branched, often with a stipe and a rather open head of branches, smooth, clear brown below, paler near the apex, with the ends of the branches often colorless. Conidia ellipsoid or obovoid, often with a slightly protruding hilum, colorless to pale brown, smooth, 6 to 18 x 4 to 4-11 μ (mostly 8 to 14 x 6-9 μ). Attacks many species of plant (121).

Economic Importance

Gray-mold blight is a weak parasite on members of the Papaveraceae and is only important during cool, damp weather when plants are nearing bloom.

Powdery Mildew

Two species of *Erysiphe* are of importance on Papaveraceae.

Taxonomy

Erysiphe cichoracearum DC. ex Merat, 1821. (Kapoor, 1967).

Erysiphe compositarum Duby, 1830.

Erysiphe scorzonerae Cast., 1845.

Erysiphe lamprocarpa Lev., 1851.

Erysiphe martii Lev., 1851.

Erysiphe tabaci Saw., 1928 (500).

Distribution and Hosts

Occurs in Europe, Formosa, Japan, and the United States mostly on *P. somniferum* but occasionally on *Chelidonium*, *Eschscholtzia*, *Meconopsis*, and a few other species of *Papaver*.

Symptoms

Mycelium and conidiophores form a white deposit on adaxial and abaxial leaf surfaces, stems, and capsules. Dark cleistothecia present in late summer and fall on infected areas.

Description

Conidia in long chains, ellipsoid to barrel-shaped, variable in size, 25 to 36 x 13 to 19 μ ; cleistothecia cinnamon brown, 95 to 151 μ ; asci 38 to 60 x 25 to 28 μ . Appendages of fruiting bodies basally inserted mycelioid, interwoven with mycelium, hyaline to dark brown, 1 to 4 times as long as diameter of ascocarp, rarely branched. Asci 10 to 15/cleistothecium; ascospores 2 per ascus 20 to 30 x 12 to 18 μ (63, 157).

Economic Importance

Powdery mildew is not too important in the field in most areas although it is destructive in the greenhouse if controls are not applied.

Control

New mildewcides are effective applied at manufacturer's directions.

Taxonomy

Erysiphe polygoni (DC.) Salm., 1900.

Distribution and Hosts

Hosts similar to those for above species. Occurs in Far East, Middle East, South America, and the United States.

Symptoms

White powdery spots on aerial parts of host (128, 609). Stunting and distortion of leaves occurs with some necrosis of invaded tissues. Plants attacked at all stages of maturity (482).

Description

Amphigenous, mycelium effuse and arachnoid; cleistothecia small, 65 to 180 μ ; appendages variable in number and length, brown or hyaline. Asci 4 to 8 spherical to pear-shaped, 54 to 60 x 10 to 34 μ , contain 4 to 8 elliptical ascospores 18 to 24 x 9 to 15 μ (128).

Epiphytology

Since conidia germinate rapidly, powdery mildew spreads unusually fast. Perithecia carry the fungus over the winter (128). In India the disease appears in February and continues until April. It develops between 20° and 30° C and is favored by shade. It tolerates low humidities and conidia will germinate at humidities from 0 to 100 percent. Internal water content of conidia is high. Light and darkness did not affect germination (262). It is particularly prevalent at higher elevations in India during damp, hot weather (78).

Economic Importance

E. polygoni is a serious parasite of opium poppy in India. Considerable loss is regularly suffered there in seed and narcotic yield (257).

Control

Destruction of weed hosts and application of chemical sprays (257, 262). One percent Solbar, a sulfur fungicide, was suggested by Flachs (128) before the advent of the newer mildewcides.

Taxonomy

- Alternaria brassicae* (Berk.) Sacc., 1880.
- Macrosporium brassicae* Berk. in Hooker, 1836.
- Alternaria brassicae* (Berk.) Sacc. var. *somniferi* Briard & Har. 1891 (53)
- Macrosporium papaveris* Bres., 1915.
- Macrosporium somniferi* Garbowski, 1918.
- Macrosporium papaveris* Parisi, 1921.
- Macrosporium bresadolae* Parisi, 1924.
- Alternaria somniferi* (Briard & Har.) Saw., 1958 (503).
- Alternaria papaveris-somniferi* Saw., 1959 (504).
- Alternaria brunnea* Saw., 1959 (504).

Distribution and Hosts

Chiefly on *P. somniferum* in Europe, Japan, and Taiwan.

Symptoms

Spotting of capsules and black; velvety-moldy lesions on leaves and flowers.

Description

Hyphae short, torulose, fasciculate, dense, 30 to 40 x 6 to 7 μ , brown, 1 to 2 septate; conidia oblong-clavate, tip subacute, 5 to 9 septate, constricted, more or less divided into compartments by longitudinal septae. Pale olivaceous, 52 to 80 x 14 to 20 μ , long-pedicellate (53).

Economic Importance

Mildly pathogenic and in general does not cause much damage. Metabolic products from this fungus have been reported to cause precocious germination of seed of *P. somniferum* at Greifswald, Germany (166).

Leaf Smut

Taxonomy

Entyloma fuscum Schroet., 1877.

Ref.: Beitr. Biol. Pflanz. (Cohn) 2: 349-385.

Sny.: *Entyloma fuscellum* Rab., 1878.

Entyloma bicolor Zopf.

Entyloma bicolor Rab., 1878.

Entyloma glaucii Dang., 1894.

Distribution and Hosts

Nearly worldwide in occurrence and chiefly found upon *P. rhoeas* and *P. somniferum* but occasionally reported upon other species of *Papaver*.

Symptoms

Pale spots (3 x 6 to 10 mm) appear on the leaves in July and August in Europe. The spots become brown and then black (128). A yellow faded area surrounds the blemish on leaves of *P. somniferum* (82).

Description

Chlamydospores (2 to 6 x 11 to 23 μ) form within the tissues within a 2 to 6 μ thick light-brown membrane. They germinate in place and a promycelium emerges through stomata. On the promycelia hyaline, fusiform sporidia (20 to 22 x 3 μ) are formed (128). Chalydospores are maroon-brown (82). No clean-cut morphological differences could be seen when the fungus was compared on *P. rhoeas*, *P. dubium*, and *P. somniferum* (82).

Epiphytology

Sporidia are readily disseminated by wind and splashing rain (82, 128).

Economic Importance

Leaf smut is not seen too frequently in Europe, therefore is not of much economic importance there (82). A problem in Hungary (128), leaf smut is also considered an important disease in Argentina (315).

Control

Crop rotation has been recommended (128).

Miscellaneous Fungi Reported upon Papaveraceae

In addition to the more important fungi that attack members of the Papaveraceae, there are many species that are of relatively minor importance upon these hosts and a number of saprophytes have also been found on them. Some of the latter have been observed on the latex. These miscellaneous fungi are listed in Appendix B with the pertinent references to them in the bibliography.

BACTERIAL DISEASES OF PAPAVERACEAE

Bacterial Blight of Opium Poppy

Taxonomy

Erwinia aroidea (Town.) Holland, 1920. Jour. Bact. 5: 222.

Syn.: *Bacillus aroidea* Townsend, 1904. U.S.D.A., Bur. Plant Ind. Bul. 60: 40 (117).

Bacterium aroideae Stapp, 1928.

Pectobacterium aroideae Waldee, 1945.

Probably syn.:

Erwinia croci (Mizusawa) Magrou, 1921.

Bacillus croci Mizusawa, 1923.

Erwinia melonis (Giddings) Holland, 1920.

Bacillus melonis Giddings, 1910.

Pectobacterium melonis Waldee, 1945.

Erwinia papaveris Christoff (Khristov), 1932.

Bacterium papaverum (Khristov) Christoff Burgv., 1935.

Erwinia papaveris (Ram Ayyar) Magrou.

Bacillus papaveris Ram Ayyar (175, 438).

Bacterium papaveris (Ram Ayyar) Burgv. 1935 (78, 117, 175).

Bacillus (*Erwinia*) *papaveris* Khristov, 1933 (237).

Distribution

Europe and Asia.

Symptoms

Dark-brown to black-brown spots on aerial parts (237) with a purplish tinge to lesions on *P. alpinum* and on *P. orientale* (21). Blackens stem and midribs of leaves in early stages; complete rotting in later stages; exudate from lesions viscous (175). When severe, leaves become dry (237).

Description

Aerobic rods 0.4 to 0.6 μ x 1 to 3.3 μ ; gram negative; 1 to 10 peritrichous flagella; single, frequently in pairs, rarely in filaments; optimum growth at 29° C, min. 6 and none at 50°; gas not produced from sugars; liquefies gelatin; no indole; acid from dextrose and sucrose (Bergey's Manual, 1957) (21); milk clears slowly without coagulation and turns brown (237). *E. aroidea* shows both pectin glycosidase and pectin methyl esterase activity (528). No qualitative differences were found in the pectic enzymes produced by the soft rot bacteria (529). Separated from *E. carotovora* to which it is closely related by serological reactions and action on maltose and xylose. *E. aroidea* does not form gas on these.

Economic Importance

Bacterial blight caused enough damage to attract attention in Formosa in 1940 (311). Regarded as of economic interest in Bulgaria (21, 237) and in India (71, 78).

Epiphytology

The bacteria are viable on seeds for 20 months (21). They enter through the stomata and are seedborne (128). Spread is rapid (237). First symptoms appear at start of bud formation. Biting and sucking insects are instrumental in transmission (461).

Control

Sanitation, crop rotation, and timely application of insecticides to prevent dissemination (128, 226, 461).

Stalk Bacteriosis of Poppy or Soft Rot

Taxonomy

Erwinia carotovora (Jones) Holl., 1920.

Syn.: *Bacillus carotovorus* Jones, 1901.

Bacterium carotovorum Lehman & Newman, 1927.

Pectobacterium carotovorum Waldee, 1945.

Distribution

Probably cosmopolitan but few reports on poppy. Europe (354).

Symptoms

Bacterial blight may appear in rosette stage in May. Aerial fleshy plant parts including leaf veins become violet. If infection occurs at bud stage, tips wilt in 2 to 3 days. Plants infected at an early stage decompose more rapidly than those infected at a later growth stage. Finally plants collapse and become brown and dry (354). Older plants may not collapse but die prematurely.

The most striking signs of stem bacteriosis are the flaccid, drooping leaves. The pith turns black-violet and is partly consumed (354).

Description

Rods 0.7 to 0.8 x 1.5 to 5.0 μ ; occasionally in filaments, peritrichous flagella; gram negative; gelatin liquefied; no indole; no H_2S ; acid and gas from sugar media; facultative anaerobe; opt. 25° to 30°, minimum 4°, maximum 38° to 39° C, thermal death point 51° C (Bergey's 1957, 7th ed, p. 355-356).

Economic Importance

Bacterial blight has caused losses in Europe in some seasons (354).

Epiphytology

Transmitted by the hop bug (*Calocoris norvegicus* (Gmll.)). Disease spreads in all directions from infection site. Favored by high field temperatures (354).

Leaf Spot Bacteriosis of Poppy

Taxonomy

Xanthomonas papavericola (Bryan and McWhorter) Dowson, 1939 (65).

Syn.: *Bacterium papavericola* Bryan and McWhorter, 1930 (65).

Phytomonas papavericola Bergey et al., 1957.

Distribution

Europe and Asia. Found on *Meconopsis argemone*, *M. bayleii*, *P. orientale*, *P. rhoeas*, and *P. somniferum*.

Symptoms

Irregular, light-colored water-soaked spots that later become yellowish and finally transparent; spots are 1 to 6 x 4 mm surrounded by necrotic zone on aerial parts. Isolated spots are larger, up to 10 mm. If numerous, the spots may merge and the leaf dries. May defoliate with abundant precipitation and warmth. If severe, may kill plants. Can become systemic by entering vascular tissue through hydathodes (516). Black lesions form on aerial parts of *P. rhoeas* and *P. orientale* (562). Seedborne if systemic (236). In *P. somniferum* infection is restricted to the leaves (461).

Description

Rods 0.6 to 0.7 x 1.0 to 1.9 μ ; filaments; monotrichous; encapsulated, gram negative; indole negative; acid but no gas from levulose, glucose, galactose, fructose, sucrose, lactose, maltose, glycerol, and mannitol; gelatin liquefied; H₂S formed; agar colonies yellow; optimum temperatures 25° to 30°, maximum 35° C (Bergey's Manual, 1957); opt. pH 6.9 to 7.6; viable 20 months in dried material (65). Thermal death point 52 (116, 516). Cross agglutination reactions place it in the "vascularum group of the Xanthomonads (122). Nutrition (528, 529, 536).

Economic Importance

Leaf spot resulted in premature defoliation in Romania in the humid 1964 season (516). Considerable damage sometimes caused by it in Bulgaria (236). With sufficient moisture and temperature the whole stand may be affected and yield is reduced (361).

Epiphytology

Bacteria are splashed from the soil to lower leaves by rain or are carried on wind-blown soil particles to leaves. Further spread is effected by plant contact, showers, and insects (461). Bacteria enter through stomata and hydathodes. First symptoms appear in early June in Romania. The attack intensifies in years when precipitation is abundant in June and July, e.g., in 1964 it caused premature defoliation. First infections are through stomata. If bacteria enter the vascular system, the disease becomes systemic. The bacteria are seedborne (236, 516). Bacteria may remain viable in plant debris as long as 20 months (516). When bacteria were injected into the stem some necrosis occurred within a few centimeter of the rupture but a soft rot was never observed (461).

Control

Sanitation, rotation, and application of protective foliar antibiotics (516).

A Bacterial Disease of Shirley Poppy

Taxonomy

Pseudomonas papaveris Lelliott and Wallace, 1955.

Ref.: Lelliott, R. A., and Maud M. Wallace, 1955 (278).

Distribution

Tanganyika (278) and India (523).

Symptoms

Similar to those incited by *Erwinia aroidea*. Sometimes first symptom is purplish-black stem discoloration at or near soil level, turning black and spreading upwards to branch tips, involving either the whole stem or initially one side only. Foliage wilts, turns yellow and then brown with veins appearing as black lines, and finally disintegrates into a malodorous slimy mass. Alternatively, black spots appear in leaf veins or blades or on the lateral branches well above soil level, either spreading to kill the plant or remaining localized. Plants are attacked at all ages, well-grown plants being killed outright in a week or two or even within a few days. Symptoms appeared within 48 hours after inoculation (278).

Description

The pathogen belongs in the green-fluorescent species of the genus. Non-lipolytic, gram negative, rod $2 \times 0.4 \mu$ average; aerobic; motile by 1 to several flagella. Gelatin not liquefied; neither nitrite, indole, nor H_2S produced, starch partly hydrolyzed; acid from xylose, dextrose, mannose, glycerol, maltose, and lactose but not from rhamnose, sucrose, raffinose, or salicin. Litmus is reduced. Optimum temperature 25° to 30° C; maximum 35° , thermal death point 49° to 50° C after 10 minutes (278).

Economic Importance

Causes damage in plantings of Shirley and oriental poppy in India (523) and of Shirley poppy in Tanganyika (278).

Epiphytology

Bacterial disease appears in plantings in India in February (523) and in Tanganyika in July, August, and September (278). Insects could be a factor in spread.

Control

Antibiotics, streptomycin, and agrimycin inhibited growth of the pathogen in vitro but were not field tested (523).

VIRUS DISEASES OF PAPAVERACEAE

Anemone Mosaci Virus (192)

Similar to viruses placed in the turnip group and serologically related to the cabbage black ringspot virus, although anemone mosaic virus infection did not protect against infection with cabbage black ringspot virus.

Distribution

Of 90 plant species, 47 were experimentally infected in England (192).

Symptoms

On *P. somniferum* slight loss of bloom and reduction of vigor at 18° C; severe dwarfing and faint systemic dark brown necrotic specks, streaks, and rings on larger leaves, when grown at 8° to 10° C. Flowers severely distorted, bleached, and often showing color breaking. *P. rhoeas* was usually a symptomless carrier but a few plants showed slight leaf chlorosis (192).

Transmission

Mechanically and by four of six aphid species tested (192).

Characteristics of the Virus

Thermal inactivation below 62° C; at 18° C inactivated in less than 72 hours; dilution end-point less than 1/2,500 (192).

Economic Importance

For *P. somniferum* damage appreciable when host at 10° C and below.

Aster Yellows (266, 267)

P. nudicaule experimentally infected was severely stunted and chlorotic. The leaf-hopper, *Cicadula sexnotata* (Fall.) transmitted the causal agent from aster (267).

Sugar Beet Yellows (29)

Sugar beet yellows was experimentally transmitted to *Eschscholtzia californica* Cham. in the United States (29) and to *P. somniferum* in Europe (83). Infected plants showed yellowing and stunting and reduced seed yield. It was transmitted by *Myzus persicae*. Discrepancies in results of tests on beet yellows virus in opium poppy may be due in part to differences in clones of *M. persicae* (464).

Active strains can eliminate seed production in poppies in the greenhouse. In the field seed yield was reduced 20 percent and seed quality suffered (83). It is possible that a 50 percent decrease in yield of poppy seed in the Netherlands was caused by a strain of sugar beet yellows virus (291). Proper timing in application of systemic insecticides to kill the vector can largely obviate the damage (83).

Cabbage Black Ring-spot Virus (322)

Occurs in South Africa on *P. nudicaule*, *P. rhoeas*, and *P. somniferum*. Mild in crucifers (cape forget-me-not and endive); moderately severe in stock (*Matthiola incana*); severe in *P. rhoeas* and *P. nudicaule*.

A severe lethal disease is developed in *P. somniferum*. It is the most sensitive of natural hosts to the disease. A short chlorotic phase may mark the start of infection, with one or two immature leaves showing vein-clearing and some chlorotic spots along the veins. This is followed almost immediately by a lethal systemic necrosis of the younger leaves. The necrosis starts with a blackening of the fine veins and the appearance of some purplish black rings towards the ends of the lobes, outside the zone of blackened veins. Interveinal areas turn black and become dry as though scorched by fire, and the leaf collapses. With the rapid spread of necrosis throughout the plant, the disease is soon fatal. Experimentally infected plants usually succumbed within 3 to 4 weeks (322).

Local lesions formed on *Nicotiana glutinosa*, *N. sylvestris*, and *N. tabacum*. Growth was not seriously checked and infection was mild. *Datura stramonium* was mostly resistant but one collection was susceptible. *Datura metel* was susceptible; petunia exhibited a systemic reaction--no local lesions; spinach developed mild to severe systemic infection. *Amaranthus tricolor* and *A. caudatus* had local necrotic lesions, no systemic infection. Wallflower (*Cheiranthus cheiri*) was systemically diseased. Radish (*Raphanus sativus*) had localized infections (322).

Mechanical transmission was successful and *Brevicoryne brassicae* and *Myzus persicae* (Sulz.) were nonpersistent vectors (173).

The virus was viable in expressed sap at 20° to 25° C for 24 to 48 hours; tolerant of 1/1,000 dilution (113); thermal inactivation point (10') was near 60° C (173). Cabbage black ring-spot virus resembles turnip virus 1 and turnip mosaic virus (173).

This virus is lethal to *P. somniferum* and causes a serious disease in *P. nudicaule* and *P. rhoeas*. It is moderately severe in stock (*Matthiola incana*). Young turnip plants are sometimes killed and older plants are much stunted. On other hosts tested little damage resulted (173).

Datura 437 Virus (93A)

In March 1969 Dr. Kahn at the Plant Introduction Station, Glen Dale, Md., isolated a virus from a tree *Datura* from Colombia. Tobacco and *P. somniferum* were among the susceptible hosts tried. *P. orientale*, *P. nudicaule*, and *Eschscholtzia californica* were not susceptible.

On *P. somniferum* chlorosis and stunting were followed by rapid death of the plants at temperatures of 25° to 30° C. Only 2 entries out of 84 representing 58 species from the World Tobacco Collection were resistant (93A).

The virus belongs in the potato Y virus group based upon particle size and length, pinwheel inclusions, and transmission characteristics. It was transmitted mechanically and inefficiently by *Myzus persicae* (Sulz.). It was not transmitted through seed, soil, or dodder. The particles are long, flexuous rods averaging 720-729 nm. In plant sap TIP is 56 to 58° C, DEP 10^{-3} to 10^{-4} , and LIV of 3 days at room temperature. It has been stored for 5 years in LN₂ in infected leaf pieces without loss of virulence.

Mosaic of Opium Poppy (Beet Mosaic Virus) (460, 506, 507)

Found in Europe on *P. somniferum* (507, 354); *Argemone intermedia* Sweet., *A. mexicana* L., *A. platyceras* Link and Otto, and *P. rhoeas* L. (506).

Diseased plants are retarded; internodes shortened near the top and leaves crowded. Leaves at first show mottling; later yellowing. Eventually only veins and adjacent tissue remain green. The stiff leaves impart a broom-shaped appearance to the plant. Capsules are almost uniformly high and are smaller than healthy ones. Seed fail to develop in the smaller capsules and become reddish-brown dust. In plants infected late in the season, seed germinability is reduced. The beet mosaic virus is not seed-borne (354).

Doralis fabae (Scop.) and *Myzus persicae* (Sulz.) are vectors (179).

Control measures include sanitation, crop rotation, and control of insect vectors (139).

Pelargonium Leaf Curl Virus (193)

This virus multiplied symptomlessly in *P. somniferum*.

Bean Yellow Mosaic Virus (*Phaseolus* Virus 2) (263, 264)

Found on *P. somniferum* in Bulgaria (263, 264). Chlorotic areas on leaves first as irregular bands along veins, later spreading often to all of interveinal areas leaving green stripes along veins; stunting. Fruits show green areas of normal turgor mixed with yellow sunken portions and yield little seed (263, 264).

Transmitted mechanically and by *Doralis* (Aphis) *fabae* (Scop.) from bean and broad bean to poppy and vice versa. Causes reduction in seed yield (263, 264).

Yellowing Virus of Poppy (460)

Found in Europe on *P. somniferum* (139, 354, 460). Leaf edges are yellow; lamina green; plants are stunted. Peduncles are elongated and often wavy; capsules are long and lopsided. Capsule partitions show yellow green to bright yellow striping, often accompanied by red-brown flecks. Seed densely crowded on swollen placentae, sometimes causing capsule walls to split. Large capsules may produce a small yield of inferior quality seed; small late capsules are barren. The dried, shriveled, and rust-colored seed are tightly bound to placentae. The reddish-brown seed of lightly infected plants are easily distinguished from healthy seed (139, 354). The disease appears annually and sporadically in all poppy fields in Europe (354).

NEMATODES REPORTED UPON POPPY

The nematodes associated with Papaveraceae have not been studied adequately. Prior to Zyubin's (622) checklist of 55 species found in Kirgizia, the following species were reported:

Heterodera^{3/} *marioni* on *P. spp.* in the United States (68); *P. orientale* in Italy (74); *P. nudicaule* in Australia (378); *P. somniferum* in Israel (448).

Heterodera^{3/} *radicicola* on *P. rhoeas* in the United States (36).

Heterodera schachtii Schmidt on *P. rhoeas* in the United States (88); on *P. somniferum* in Czechoslovakia (512) and in Germany (242).

Meloidogyne sp. on *P. somniferum* in Israel (335).

Meloidogyne arenaria on *P. rhoeas* and *P. nudicaule* in Rhodesia and Nyasaland (302).

^{3/} These species now included in *Meloidogyne*.

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(469)

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(470)

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(471)

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(474)

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(476)

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P. rhoeas; *Erysiphe polygoni*, *Meloidogyne* sp., *Rhizoctonia solani*, and *Xanthomonas papavericola* on *P. somniferum*; *Entyloma fuscum*, *Septoria* sp., and *Xanthomonas papavericola* on *P. spp.*; *Phymatotrichum omnivorum* and *Septoria chelidonii* on *Chelidonium majus*, pp. 328-329.

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451 V67.

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1953. The plant diseases of Nyasaland. Commonwealth Mycol. Inst.
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Phytophthora parasitica on *Eschscholtzia californica*, p. 16.

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1908. Studies in North American Peronosporales. III. New or
noteworthy species. Torrey Bot. Club Bul. 35: 361-365.
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on *Argemone platyceras*.

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1887. Ascomyceten [Ascomycetes.] Rabenhorst's Kryptogamen Flora.
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Pleospora papaveracea on *P. somniferum*, p. 501.

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- (609) Zaman, M. B., and Khan, A. S.
1969. Studies on some diseases of *Papaver somniferum* Linn. Pakistan Jour. Forest. 19: 185-193. 99.8 P17.
Erysiphe polygoni, *Cercospora papaveri*, *Rhizoctonia solani*, *Peronospora arborescens*, *Dactylium roseum*, *Trichoderma viride*, *Sporotrichum* sp., *Cladosporium herbarum* and *Rhizomorpha* sp.
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Pleospora calvescens [*Helminthosporium papaveris*]
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1925. Diseases of cultivated plants in Middle Asia. [Transl. from Russ.] 165 pp. Tashkent, Uzbek SSR. 464 Z12. RAM 5: 174-175.
Peronospora arborescens (Uzbek SSR) and *Erysiphe cichoracearum* (Turkistan) on *P. somniferum*, p. 15.

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1958. Mikoflora nasion maku. [The fungous flora of opium poppy seeds.] Rocz. Nauk, Rolnicz. Ser. A, — Roslinna 78: 309-342, illus. 20.5 R59 Ser. A.
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Cladosporium herbarum, *Fusarium concolor*, *F. scripi* var.
caudatum, *Gloeosporium* sp., *Helminthosporium papaveris*,
Penicillium spp., *Pullularia pululans*, *Rhizopus nigricans*,
Trichoderma sp., *Trichothecium roseum*, and yeast.
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[First list of fungi of Mexico. Arranged by hosts.] Mex.
Ofic. de Estud. Espec. Fol. Tec. 14, 43 pp. 8 M577F.
- Phymatotrichum omnivorum* on *Argemone* sp., p. 15.
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- On *P. rhoeas*.
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1944. Die Herzfäule des Oelmohns und ihre Bekämpfung. [The heart rot of the oil poppy and its control.] Switz. Eidg. Landw. Versuchsanst. Zurich-Oerlikon, Flugbl. 14: 1-4. 105.5 Oe7.
- Boron deficiency.
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1945. Die Blattdürre des Mohns. [Poppy leaf blight.] Schweiz. Naturf. Gesell., Verhandl. 125: 172. 508 B452. RAM 26: 126.
- Dendryphium penicillatum* [*Pyrenophora calvescens*]
- (617)

1945. Die Blattdürre des Mohns (*Pyrenophora calvescens* [Fr.] Sacc. nebenfruchtform *Dendryphium penicillatum* [Cda.] Fr.) [Poppy leaf blight (*Pyrenophora calvescens*, perfect stage of *Dendryphium penicillatum*).] Schweiz. Bot. Gesell. Ber. 55: 240-269, illus. 451 Sch9.

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1946. Zur Kenntnis Pflanzlichen Abwehrreaktionen. Der Einfluss der Temperatur auf das Zustandekommen der gummösen Demarkationszone. [Contributions to the knowledge of plant defense reactions. The influence of temperature on the development of the gummy demarcation zone.] Schweiz. Bot. Gesell., Ber. 56: 507-522. 451 Sch9.

Dendryphium penicillatum [*Pyrenophora calvescens*] on *P. somniferum*.

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Entyloma fuscum on *P. somniferum* and *P. rhoeas*, p. 1025.

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Entyloma fuscum on *P. rhoeas*, p. 167, So. Africa.

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1962. Protection of opium poppy from pests and diseases. [Transl. from Russ.] Sel'sk. Khoz. Kirgizii 1962 (1): 31-33.

Peronospora.

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1969. Plant nematodes of the opium poppy in Kirgizia. (Transl. from Russ.) 100 pp. Akad. Nauk Kirgizskoi SSR., Frunze.

The following nematodes have been reported on *P. somniferum*:

Acrobeloides buetschlii=*Cephalobus persegneis*=*Acrobeles buetschlii*=*Cephalobus persegneis* f. *buetschlii*
Acrobeloides uberrimus
Aglenchus costatus=*Tylenchus elegans*=*T. buffalora*
Alaimus primitivus
Aphelenchus avenae=*Aphelenchus agricola*=*Aphelenchus maupasi*
Aphelenchoides fragariae=*Aphelenchus fragariae*=
Aphelenchoides olesistus=*A. (Chitinoaphelenchus) olesistus*=
Aphelenchus olesistus f. *longicollis*

Aphelenchoides parietinus=*Aphelenchus parietinus*=
A. striatus f. *aquaticus*=*A. aquatica*=*Cephalobus alpinus*=
Pathoaphelenchus parietinus=*Tylenchus bulbosis*
Aphelenchoides saprophilus
Aphelenchoides subparietinus
Aphelenchoides subtenuis=*Aphelenchus subtenuis*=
Aphelenchoides hodsoni
Cephalobus persegnis
Chiloplacus symmetricus=*Acrobeles* (*Acrobeloides*)
symmetricus=*Cephalobus symmetricus*
Diphtherophora communis
Ditylenchus destructor
Ditylenchus dipsaci=*Anguillula dipsaci*=*A. devastatrix*=
A. secale=*A. putrefacicus*
Dorylaimus filiformis=*D. polyblastus*=*D. zografii*=
D. stagnalis filiformis=*D. filiformis* f. *salinae*
Doryllium uniforme
Eucephalobus elongatus=*Cephalobus elongatus*=*Neocephalobus*
elongatus=*Cephalobus* (*Eucephalobus*) *elongatus*
Eucephalobus oxyuroides=*Cephalobus oxyuroides*
Eucephalobus striatus=*Cephalobus bursifer*
Eudorylaimus acuticauda=*Dorylaimus acuticauda*
Eudorylaimus obtusicaudatus
Eudorylaimus papillatus=*Dorylaimus papillatus*=
D. domus-glauci
Eudorylaimus paraobtusicaudatus=*D. samarcandicus*
Filenchus filiformis=*Tylenchus elegans*=*T. exiguus*=
T. pillulifer
Helicotylenchus multicinctus
Helicotylenchus nannus
Heterodera schachtii
Hexatylus viviparus=*Jotonchium viviparum*
Lelenchus infirmus=*Anguillulina leptosoma* f. *minuta*
Mesodiplogaster lheritieri=*Diplogaster lheritieri*=
Fristionchus lheritieri
Mesodorylaimus bastiani=*Dorylaimus bastiana*=*D. landii*=
D. javanicus=*D. intermedium*=*D. tenuicaudatus*
Monhystera agilis
Monhystera bulbifera
Mononchus papillatus
Nothotylenchus acris
Panagrolaimus hygrophilus=*Panagrolaimus thienemanni*
Panagrolaimus rigidus=*Leptodera rigida*=*Anguillula rigida*=
Cephalobus rigidus=*Cephalobus oxyurus*=*Rhabditis aquatica*
Panagrolaimus subelongatus=*Cephalobus subelongatus*
Paraphelenchus pseudoparietinus=*Aphelenchus*
(Paraphelenchus) pseudoparietinus
Paratylenchus hamatus

Paraphelenchus tritici
Plectus granulatus=*P. schneideri*=*P. blanci*=*P. tubifer*
Pratylenchus montanus
Pratylenchus scribneri=*Tylenchus penetrans*
Pratylenchus vulnus=*Anguillulina pratensis*=*Pratylenchus*
musicola
Prismatolaimus dolichurus
Prismatolaimus intermedius=*Monhystera intermedia*=
Prismatolaimus hawaiiensis
Rhabditis brevispina=*Anguillula brevispina*=*Leptodera*
cucumeris
Rhabditis intermedia
Tetylenchus productus
Tylenchorhynchus dubius=*Anguillulina dubia*
Tylenchus agricola=*Tylenchus filiformis*
Tylenchus davainei

Fungi

Aecidium plenum Arth., 1918.

Texas 580.

Alternaria spp.

Syn.: *Macrosporium* spp.

Denmark 372.

Israel 448.

Italy 415.

Poland 334, 612.

Texas 580.

Alternaria alternata (Fr.) Keissl., 1912.

Syn.: *A. tenuis* Nees 1816/17.

Morocco 452.

Poland 328.

Alternaria brassicae (Berk.) Sacc., 1880.

Syn.: *A. brassicae* (Berk.) Sacc. var. *somniferi* Briard & Har., 1891.

A. brunnea Saw., 1959.

A. papaveris-somniferi Saw., 1959.

A. somniferi (Briard & Har.) Saw., 1958.

Macrosporium bresadolae Parisi, 1924.

M. commune Rab. 1870.

M. papaveris Parisi, 1921.

M. somniferi Garbowski, 1918.

Bulgaria 239.

Czechoslovakia 512.

France 53, 474.

Germany 52, 166.

Italy 396, 397.

Japan 503.

Latvian SSR. 514, 526.

Poland 141.

Sakhalin, USSR. (formerly Japanese) 205, 206, 207.

Taiwan 504.

Ukrainian SSR. 140.

Alternaria brassicae (Berk.) Sacc. var. *somniferi* Briard & Har., 1891.

= *A. brassicae* (Berk.) Sacc., 1880.

^{5/} Numbers after geographical locations refer to entries in the bibliography.

Alternaria brunnea Saw., 1959. = *A. brassicae* (Berk.) Sacc., 1880.

Alternaria lancipes Ell. & Ev., 1888.

Kansas 120, 474.

Texas 580.

USA 126.

Alternaria papaveris-somniferi Saw. = *A. brassicae* (Berk.) Sacc., 1880.

Alternaria somniferi (Briard & Har.) Saw. = *A. brassicae* (Berk.) Sacc., 1880.

Alternaria tenuis Nees, 1816/17. = *A. alternate* (Fr.) Keissl., 1912.

Alternaria tenuissima (Nees ex Fr.) Wilts., 1966.

Sudan 552.

Aregma moniliforme = *Phragmidium mucronatum* (Pers.) Schlecht.

Ascochyta sp.

Lithuanian SSR 62.

Ascochyta chelidonii Kab. & Bub., 1907.

Czechoslovakia 480.

Ascochyta papaveris Oud., 1885.

Syn.: *Diplodinia papaveris* (Oud.) Lind, 1926.

Novaya Zemlya, USSR 474.

South Africa 109.

Spitsbergen 280, 281.

Aspergillus spp. (sapro.)

Germany 449.

Poland 612.

Aureobasidium pullulans (de By.) Arnaud., 1918.

Syn.: *Pullularia pullulans* (de By.) Berk.

Poland 612.

Botryotinia fuckeliana (de By.) Whet., 1945. (Imperfect stage is *Botrytis cinerea* Pers. ex Fr. 1801.)

Alaska 13, 580.
Japan 138.
Maryland 580.
Moscow Oblast 429.
New Zealand 54.
Poland 141.
Spitsbergen 280.
Wisconsin 580.

Botrytis spp.

Germany 450.
Sweden 450.

Botrytis arborescens Berk., 1846 = *Peronospora arborescens* (Berk.) de By., 1863.

Botrytis cinerea Pers. ex Fr., 1801 = *Botryotinia fuckeliana* (de By.) Whet., 1945.

Caeoma chelidonii Magn., 1875.

Belgium 421.
France 421.
Germany 471.
New York 471.
Switzerland 421.

Cercospora sp.

Venezuela 531.

Cercospora papaveri Mueller & Chupp, 1936 = *C. papavericola* Chupp, 1954.

Cercospora papaveri Nakata = *C. papavericola* Chupp.

Cercospora papavericola Chupp

Syn.: *C. papaveri* Mueller & Chupp
C. papaveris Saw.
C. papaveri Nakata

Alabama 81, 538, 580.
Brazil 81, 356.
Florida 81.
India 401, 402.
Japan 138, 606.
Kazakhstan, SSSR 225.
Texas 81.
Taiwan 81, 504.
West Pakistan 233, 609.

Cercospora papaveris Saw. = *C. papavericola* Chupp

Cercospora whetzelii Chupp in Toro, 1931.

Puerto Rico 81, 569, 580.
Venezuela 81.

Cercosporidium guanicense (F. L. Stevens) Deighton, 1967.
Syn.: *Cladosporium guanicense* F. L. Stevens (sapro.)

Bermuda 597.
Brazil 539.
Cuba 539.
Dominican Republic 539.
Puerto Rico 537, 539.
Venezuela 531, 539.

Chaetomella atra Fckl.

Greece 10, 392.

Cladosporium sp.

Sudan 552.

Cladosporium cladosporioides (Fres.) de Vries

Sudan 552.

Cladosporium guanicense F. L. Stevens = *Cercosporidium guanicense*
(F. L. Stevens) Deighton, 1967.

Cladosporium herbarum (Pers.) Link ex. Gray, 1921. (sapro.)

Czechoslovakia 428.
Germany 449.
India 20.
Karelian ASSR 277.
Kazakhstan, USSR 225.
Kirghiz SSR 422.
Kirov Oblast 135.
Poland 141, 328, 612.
Tomsk, Siberia 276.
Turkmen SSR 223, 256.
West Pakistan 609.

Clathrospora pentamera (Karst.) Berl.

Spitsbergen 280, 281.

Cucurbitaria papaveracea de N. = *Pleospora papaveracea* (de N.) Wint., 1887.

Curvularia geniculata (Tracy & Earle) Boed.

Turkey 154.

Dacrymyces papaveris Quel.

France 430, 475.

Dactylium roseum Berk.

India 20.

West Pakistan 609.

Dematium pullulans de By.

Victoria, Australia 321.

Queensland, Australia 321.

Dendryphium papaveris (Saw.) Saw., 1959 = *Pleospora papaveracea* (de N.) Wint., 1887.

Dendryphium penicillatum (Cda.) Fr. = *Pleospora papaveracea* (de N.) Wint., 1887.

Dendryphium penicillatum (Cda.) Fr. var. *sclerotiale* Meffert, = *Pleospora papaveracea* (de N.) Wint., 1887.

Dendryphium ramosum Cooke, 1871 = *Pleospora papaveracea* (de N.) Wint., 1887.

Didymaria chelidonii Jacz. in Komarov

Amur Oblast, USSR 254, 477.

Didymellina dianthi Burt

Syn.: *Heterosporium echinulatum* Harkn. (sapro.)

Bulgaria 239.

Diplodina chelidonii N. Naum., 1914

Latvian SSR 514.

Diplodina eschscholtziae Oudem.

Netherlands 479.

Diplodina papaveris (Ouds.) Lind, 1926 = *Ascochyta papaveris* Oud., 1885.

Diplodinula glaucii (Cke. & Mass.) Fl. Tassi

England 555.

Entyloma sp.

Lithuanian SSR 62.

Entyloma bicolor Zopf in Rab., 1878 = *E. fuscum* Schroet., 1877.

Entyloma chelidonii Cif., 1924.

Finland 285.

Entyloma eschscholtziae Harkn., 1884.

California 580.

USA 126.

Entyloma fuscellum Rab. = *E. fuscum* Schroet., 1877.

Entyloma fuscum Schroet., 1877.

Syn.: *E. bicolor* Zopf. in Rab.

E. fuscellum Rab.

Argentina 300.

Belgium 39.

Bermuda 596.

Cyperus 147, 363.

Denmark 279.

Dominican Republic 82.

Finland 285.

France 314, 316, 375, 471, 586.

Germany 288, 326, 433, 471.

Great Britain 471, 483.

Greece 18, 295, 392.

Hungary 50, 60.

Iowa 580.

Lithuanian SSR 64.

Maine 580.

Morocco 452.

Netherlands 386.

New Zealand 56, 60.

Poland 141, 226, 250.

Romania 44, 107, 490, 491, 493.

South Africa 109.

Spain 575, 577.

Switzerland 315, 317, 320.
Texas 580.
Tomsk, Siberia 276.
Turkmen SSR 223, 256.
Turkey 49, 153.
West Pakistan 8, 9.
Yugoslavia 284.

Epicoccum herbarum Croda, 1888. (sapro.)

Czechoslovakia 470.

Erysiphe spp.

Germany 274, 379.
India 519.
Poland 226.

Erysiphe cichoracearum DC. ex Merat
Syn.: *E. communis* (Wallr.) Blum.
E. tabaci Saw., 1928.

Finland 443.
France 94, 95.
Hungary 574.
India 74, 188, 398.
Japan 138.
Kazakhstan, USSR 188.
Lithuanian SSR. 63, 188.
Romania 107.
Siberia 188.
Sudan 551.
Switzerland 319, 320.
Taiwan 188, 499, 500.
Turkestan, USSR 611.
Ukrainian SSR 188.

Erysiphe cichoracearum DC. em. Salm. f. *papaveris* Pot.

Kazakhstan, USSR 224, 225.
Kirghiz SSR 422.

Erysiphe communis (Wallr.) Blum. = *E. cichoracearum* DC. ex Merat

Erysiphe communis (Wallr.) Fr. = *E. cichoracearum* DC. ex Merat

Erysiphe communis (Wallr.) Lk. = *E. cichoracearum* DC. ex Merat

Erysiphe communis Grev. = *E. martii* Lev.

Erysiphe martii Lev.

Syn.: *E. communis* Grev.

Lithuanian SSR 63.

Erysiphe polygoni DC., 1805.

Argentina 22, 176, 177.

California 432, 580, 607.

France 316.

India 72, 73, 78, 257, 260, 262.

Israel 80.

Japan 189.

Lithuanian SSR 64.

Nyasaland 602.

Oregon 517, 580.

Switzerland 37, 315.

Tasmania 7.

West Pakistan 609.

Yugoslavia 440, 441.

Erysiphe tabaci Saw., 1928 = *E. cichoracearum* DC. ex Merat

Fusarium spp.

Bulgaria 239.

Israel 448.

Poland 141.

Fusarium concolor Reinking, 1934.

Poland 612.

Fusarium equiseti (Cda.) Sacc.

Manitoba 159.

Fusarium martii Appel & Wr., 1910.

Spain 367.

Fusarium oxysporum Schl. emend. Snyder & Hans.

Manitoba 159.

Fusarium [*oxysporum* Schlecht. var.] *cubense* (E. F. Sm.) Wr., 1935.

Spain 367.

Fusarium scirpi Lamb. & Fautr.

South Africa 108, 109.

Fusarium scirpi Lamb. & Fautr. var. *caudatum* Wr., 1931.

Bulgaria 239.

Poland 612.

Fusarium scirpi Lamb. & Fautr. var. *compactum* Wr., 1931.

South Africa 108.

Fusarium solani (Mart.) Appel & Wr., 1910 emend Snyder & Hans.

Manitoba 159.

Gloeosporium spp.

Japan 180, 181, 182.

New Jersey 405.

Poland 612.

Gloeosporium argemones Ell. & Ev., 1887 = *Glomerella cingulata* (Ston.) Spauld. & Schrenk, 1903.

Glomerella cingulata (Ston.) Spauld. & Schrenk, 1903.

Syn.: *Gloeosporium argemones* Ell. & Ev., 1887.

Kansas 119, 474, 541, 580.

New Jersey 405.

Texas 580.

USA 126.

Helicobasidium purpureum (Tul.) Pat.

Great Britain 406.

Helminthosporium spp. = *Pleospora papaveracea* (de N.) Wint., 1887.

Helminthosporium papaveris E. Henning = *Pleospora papaveracea* (de N.) Wint., 1887.

Helminthosporium papaveris Saw. = *Pleospora papaveracea* (de N.) Wint., 1887.

Heteropatella lacera Fckl.

Syn.: *H. umbilicata* (Fr.) Jaap

Barnul, USSR 360.

Finland 281.

Greenland 281.

Heteropatella umbilicata (Fr.) Jaap = *H. lacera* Fckl.

Heterosporium echinulatum Harkn. (sapro.) = *Didymellina dianthi* Burt

Heterosporium eschscholtziae Harkn., 1884.

California 470, 580.

USA 126.

Heterosporium groenlandicum Allesch., 1897.

Greenland 476.

Kursk, USSR 423.

South Africa 109.

Ukrainian SSR 423.

Hypocrea spinulosa Fckl.

Finland 468.

Leptosphaeria cheldonii Fautr., 1895.

France 476.

Leptosphaeria papaveris Rostr., 1903.

Iceland 271, 478.

Leptosphaeria pellita (Rab. & Klotzsch.) Sacc.

France 466.

Leveillula papaveracearum Golov., 1956.

Uzbek SSR 188.

Leveillula papaveracearum Golov. f. *argemones* Golov., 1956.

Uzbek SSR 156.

Leveillula taurica (Lev.) Arn., 1921.

France 188.

Sudan 188, 552.

Taiwan 188.

Tanganyika 188, 453.

Uzbek SSR 188.

West Pakistan 188, 233.

Lophodermium svalbardense Lind., 1928.

Spitsbergen 280.

Macrophoma papaveris Camara, 1930.

Portugal 75.

Macrophomina phaseoli (Maubl.) Ashby = *M. phaseolina* (Tassi) Goid., 1950.

Macrophomina phaseolina (Tassi) Goid., 1950.

Syn.: *M. phaseoli* (Maubl.) Ashby, 1927.

Rhizoctonia bataticola (Taub.) Butl., 1925.

India 78, 101.

Macrosporium spp. = *Alternaria* spp.

Macrosporium bresadolae Parisi, 1924 = *Alternaria brassicae* (Berk.)
Sacc., 1880.

Macrosporium commune Rab., 1870 = *Alternaria brassicae* (Berk.) Sacc.
1880.

Macrosporium papaveris Bres., 1915 = *Alternaria brassicae* (Berk.)
Sacc., 1880.

Macrosporium papaveris Parisi, 1921 = *Alternaria brassicae* (Berk.)
Sacc., 1880.

Macrosporium somniferi Garbowski, 1918 = *Alternaria brassicae* (Berk.)
Sacc., 1880.

Massaria cleistotheca Harkn., 1834.

California 473.

USA 126.

Melampsora chelidonii-pierotii T. Matsu., 1926.

Japan 310.

Melampsora larici-populina Kleb.

Latvian SSR 514.

Melampsora magnusiana Wagner

Austria 547.

Caucasia, USSR 520.

Denmark 279, 547.

Finland 547.

Germany 547.

Hungary 547.

Japan 208.
Lithuanian SSR. 333.
Norway 547.
Sweden 388, 547.

Metasphaeria patelliformis Kirsch., 1935.

Latvian SSR 244.

Mucilago spongiosa (Leyss.) Morg. (sapro.)

Great Britain 100, 134.

Mucor mucedo Fr. (sapro.)
Syn.: *M. mucedo* (L.) Lk.

Bulgaria 239.
India 20.

Mucor mucedo (L.) Lk. = *M. mucedo* Fr. (sapro.)

Mycosphaerella arthopyrenioides (Auers.) Larsen, 1932
Syn.: *Sphaerella arthopyrenioides* Auers., 1869

Austria 467.
Iceland 271.
USA 126.

Mycosphaerella chelidonii (Fautr. & Lambotte) Tomilin, 1969.
Syn.: *Sphaerella chelidonii* Fautr. & Lambotte, 1895.

France 476.

Mycosphaerella pachyasca (Fckl.) Berl. & Vog.

Canada (northern coastline) 98.

Mycosphaerella schoenoprasi (Rab.) [not validly published.] = *Sphaerella schoenoprasi* (Rab.) Auersw.

Mycosphaerella tassiana (de N.) Johans., 1885.
Syn.: *Physalospora polaris* Rostr.
Sphaerella arthopyrenioides Auers.
Sphaerella karajacensis Allesch., 1897.
Sphaerella schoenoprasi (Rab.) Auersw.

Barnul, USSR 360.
China 413.
Ellesmere Land, N. W. Territories, Canada 281.
Norway 281.

Novaya Zemlya, USSR 281.

Spitsbergen 280.

Mycosphaerella tulasnei Jancz.

Bulgaria 239.

Myrothecium sp.

Bermuda 597.

Oidiopsis sp.

India 258.

Oidiopsis papaveris Saw., 1933.

Taiwan 501.

Oidium spp.

Armenian SRR 188.

Australia 188.

Bulgaria 188.

France 19, 188.

Great Britain 188.

India 188, 398.

Israel 188.

Italy 188.

Lithuanian SSR 63.

New Zealand 54, 188.

Portugal 188.

Southern Rhodesia 188, 195, 196, 197.

Spain 188.

Tanganyika 188, 453.

Oidium erysiphoides Fr., 1832.

Argentina 530.

Bulgaria 238.

India 73, 338, 399.

Japan 482, 502.

Oidium erysiphoides Fr. f. *papaveris* Roum.

France 456.

Ophiobolus sativus (Pam., King & Bakke) Ito & Kurib., 1929.

Bulgaria 239.

Ophiobolus tenellus (Awd.) Sacc.

Denmark 279.

France 91.

Ovularia sp.

Venezuela 531.

Ovularia indica R. Rao, 1968.

India 442.

Pellicularia filamentosa (Pat.) Rogers, 1943 = *Thanatephorus cucumeris*
(Frank) Donk, 1956.

Penicillium spp. (sapro.)

Bulgaria 239.

Poland 612.

Peronospora spp.

Kirgiz SSR 621.

Thailand 77.

Peronospora arborescens (Berk.) de By., 1863.

Syn.: *Botrytis arborescens* Berk., 1846

P. arborescens (Berk.) de By. f. *papaveris somniferi* Sacc.

P. effusa Rab. var. *papaveris* Desm.

P. effusa Rab. f. *papaveris* Fckl.

P. grisea Ung., 1929.

P. grisea minor Casp., 1857.

P. papaveris Tul.

Algeria 169.

Argentina 169, 282, 283.

Austria 169, 198, 199, 427, 508, 563.

Azerbaijan SSR 114.

Belgium 34, 87, 471.

Belorussian SSR 1.

Bulgaria 169, 239.

China 339, 387, 548, 565.

Czechoslovakia 265, 354, 407, 416, 512.

Denmark 169, 174, 279, 533, 534.

Egypt 57, 169, 327.

Estonia 66, 169.

Finland 169.

France 19, 87, 94, 95, 137, 169, 314, 316, 466, 471.

Georgian SSR 588.

Germany 26, 27, 34, 67, 87, 167, 169, 209, 213, 215, 243, 293, 294,
307, 380, 393, 431, 471.

Great Britain 25, 32, 34, 87, 89, 100, 134, 169, 406, 471.

Hungary 343, 574.
 India 20, 70, 71, 72, 73, 78, 103, 169, 257, 259, 261, 308, 338, 399,
 424, 513, 543, 561.
 Iran 169, 414, 505.
 Israel 390.
 Italy 34, 87, 169, 471, 553.
 Japan 138, 169, 183, 202, 204, 312, 546.
 Kazakhstan, USSR 225.
 Kirgiz SSR 422.
 Kirov Oblast, USSR 135.
 Kursk Oblast, USSR 42.
 Latvian SSR 40, 41, 66, 169, 514, 525.
 Leningrad Oblast 66, 366.
 Libya 296.
 Morocco 297, 452.
 Moscow Oblast 106, 429.
 Netherlands 169, 289, 385, 386, 418.
 New South Wales 3, 4, 377.
 Pakistan 169.
 Palestine 444.
 Poland 139, 141, 203, 226, 272, 417, 481.
 Portugal 158, 169.
 Pskov Oblast, USSR 286.
 Romania 44, 109, 116, 169, 383, 454, 490, 492, 494, 495.
 Sakhalin, USSR (formerly Japanese) 206.
 Spain 287.
 Stavropol', USSR 347, 361.
 Sweden 169, 171, 389, 567.
 Switzerland 169, 315, 317, 320.
 Tunisia 400.
 Turkey 48, 153.
 Turkmen SSR 223, 256.
 Ukrainian SSR 151, 410.
 United States
 Colorado 200, 603.
 Texas 580.
 Uruguay 2, 249, 346.
 Ussuri Oblast, USSR 365.
 Uzbek SSR 611.
 West Pakistan 8.
 Yugoslavia 46, 92, 137, 162, 169, 172, 216, 219, 220, 221, 248, 253,
 332, 394, 439, 441, 524, 540.

Peronospora arborescens (Berk.) de By. f. *papaveris somniferi* Sacc. = *P.*
arborescens (Berk.) de By., 1863.

Peronospora argemones Gaeum., 1923.

Czechoslovakia 169.

Denmark 169.
Esthonia 169.
France 169.
Germany 169.
Great Britain 169.
Israel 169, 390.
Kazakhstan, USSR 225.
Latvian SSR 169.
Palestine 444.
Poland 455.
Sweden 169.

Peronospora chelidonii Miy.

China 387.
Japan 191, 545, 566.

Peronospora cristatum Tranz.

Israel 390, 444.

Peronospora effusa Rab. var. *papaveris* Desm. = *P. arborescens* (Berk.)
de By., 1863.

Peronospora effusa Rab. f. *papaveris* Fckl. = *P. arborescens* (Berk.)
de By., 1863.

Peronospora gaeumanni Mayor, 1949 = *P. meconopsidis* Mayor, 1958.

Peronospora gaeumanni Mund., 1938.

India 358, 399, 403.
West Pakistan 148, 233.

Peronospora glaucii Lob.

Syn.: *P. glaucii* Savul. & Rayss, 1935.

Romania 492, 496.

Peronospora glaucii Savul. & Rayss, 1935 = *P. glaucii* Lob.

Peronospora grisea Ung., 1847 = *P. arborescens* (Berk.) de By., 1863.

Peronospora grisea minor Casp., 1857 = *Peronospora arborescens* (Berk.)
de By., 1863.

Peronospora meconopsidis Mayor, 1958.

Syn.: *P. gaeumanni* Mayor, 1949.

Switzerland 320.

Peronospora papaveris Tul. = *P. arborescens* (Berk.) de By., 1863.

Peronospora parasitica de By.

Germany 393.

Peziza eschscholtziae Phill. & Harkn., 1884.

United States 126.

Phoma melaena (Fr.) Mont. & Dur.

Hungary 51.

Phoma morphea Sacc., 1881.

Italy 465, 469.

Phoma rhoeadis Brun.

France 475.

Sweden 369.

Phoma sphaeronaemoides Fautr., 1895.

France 476.

Phoma striaeformis Dur. & Mont. var. *hysteriola* Sacc.

France 61, 466, 469.

Phragmidium mucronatum (Pers.) Schlecht.

Syn.: *Aregma moniliiforma*

India 20.

Phyllosticta chelidonii Bres., 1896.

Germany 476.

Phymatotrichum omnivorum (Shear) Dug., 1916.

California 240.

Mexico 613.

Texas 580.

Physalospora polaris Rostr. = *Mycosphaerella tassiana* (de N.) Johans.

Phytophthora spp.

Australia 572.
Victoria, Australia 58.
Western Australia 76.

Phytophthora cactorum (Leb. & Cohn) Schroet., 1886.

Netherlands 376, 572.
Scotland 100, 134.

Phytophthora cryptogea Pethyb. & Laff., 1919.

Germany 379.
New South Wales 3, 377, 378.
Tasmania 6.
Victoria, Australia 59.

Phytophthora nicotiana Breda de Haan var. *parasitica* (Dast.) Waterhouse, 1963.

Syn.: *P. parasitica* Dast., 1913.

Nyasaland 602.
Scotland 100, 134.

Phytophthora parasitica Dast., 1913 = *P. nicotiana* Breda de Haan var. *parasitica* (Dast.) Waterhouse, 1963.

Phytophthora verrucosa Alcock & Foister In Foister, 1940.

Scotland 100, 133, 134, 595.

Pleosphaerulina californica Berl.

California 476, 580.

Pleospora spp.

Canada (northern coast-line) 98.

Pleospora argyrospora Harkn., 1884.

California 473.
United States 126.

Pleospora calvescens (Fr.) Tul. = *P. papaveraceae* (de Not.) Wint., 1887.

Pleospora herbarum (Fr.) Rab., 1873.

Imperfect stage is *Stemphylium botryosum* Wallr.

Canada (northern coast-line) 93, (Cape Chudleigh) 118.
Caucasia, USSR 587.

Denmark 279.
Palestine 445.
Spitsbergen 280.
Tomsk, Siberia 276.
United States 126.

Pleospora infectoria Fckl.
Syn.: *P. vulgaris* Niessl

Canada (northern coast-line) 98.
France 91.
Spitsbergen 280.

Pleospora media Niessl.

Alaska 580.

Pleospora papaveracea (de N.) Sacc., 1883 = *P. papaveracea* (de N.)
Wint., 1887.

Pleospora papaveracea (de N.) Wint., 1887.
Syn.: *Dendryphium papaveris* (Saw.) Saw., 1959.
D. penicillatum (Cda.) Fr.
D. penicillatum (Cda.) Fr. var. *sclerotial* Meffert, 1950.
D. ramosum Cooke
Helminthosporium papaveris E. Henn.
H. papaveris Saw., 1918.
Pleospora calvescens (Fr.) Tul.
P. papaveracea (de N.) Sacc., 1883.

Armenian SSR 522.
Austria 198.
Bulgaria 234, 235.
Czechoslovakia 129, 130, 131, 132, 230, 348, 349, 350, 351, 352, 353,
354, 470, 598.
Denmark 363, 369, 371, 373, 374.
France 19, 91, 95, 112, 459, 470, 610.
Germany 23, 110, 124, 163, 164, 165, 167, 168, 307, 325, 364, 379,
380, 447, 449, 450, 451, 509, 510, 559.
Great Britain 470.
Hungary 604.
India 121.
Israel 448.
Italy 458, 468, 470, 604.
Kazakhstan, USSR 225.
Kirghiz SSR 422.
Latvian SSR 514.
Morocco 452.
Netherlands 385, 419, 420.
New South Wales 591.

Poland 24, 93, 139, 612.
Romania 107, 404, 434, 435, 486.
Sakhalin, USSR 206.
South Africa 121.
Sweden 30, 31, 115, 450.
Switzerland 616, 617, 618.
Taiwan 121, 498, 504, 550.
Tanganyika 102.
Turkey 154.
Ukrainian SSR 38, 149, 150, 152, 336.
United States 126.
Yugoslavia 248, 330, 331, 332, 381, 439.

Pleospora pellita (Fr.) Rab. = *P. papaveracea* (de N.) Wint., 1887.

Pleospora scrophulariae (Desm.) Hoehn., 1917.

Franz Josef Land, USSR 281.
Ellismere Land, Canada 281.

Pleospora vulgaris Niessl = *P. infectoria* Fckl.

Pocosphaeria dendromeconis Earle, 1904.

California 480, 580.

Pullularia pululans (de By.) Berk. = *Aureobasidium pullulans* (de By.)
Arnaud.

Pyrenophora androsaces (Fckl.) Sacc.

Spitsbergen 280.

Pyrenophora calvescens (Fr.) Sacc. = *Pleospora papaveracea* (de N.) Sacc.

Pyrenophora cerastii (Oud.) Lind, 1924.

Spitsbergen 280.

Pyrenophora chrysospora (Niessl) Sacc., 1884.

Spitsbergen 280.

Pyrenophora paucitricha (Fckl.) Berl. & Vogl.

Canada (northern coast-line) 98.

Pyrenophora pellita (Fr.) Sacc., 1883 = *Pleospora papaveracea* (de N.) Wint.

Pythium spp.

Australia (Canberra) 17.

South Africa 108.

Pythium debaryanum Hesse, 1874.

Netherlands 329.

Pythium mamillatum Meurs, 1928.

Australia (Canberra) 15, 16.

Pythium megalacanthum de By., 1881.

Japan 329.

Pythium oligandrum Drechs., 1930.

South Africa 109, 329, 590.

Pythium spinosum Saw., 1926.

Japan 329.

Pythium ultimum Trow, 1901.

Australia (Canberra) 14, 15, 16.

South Africa 109, 269, 329, 568, 590.

United States 329.

Rhabdospora rhoeadis Tassi, 1904.

Italy 479, 556, 557.

Rhizoctonia spp.

India 70, 71.

Indiana 142.

South Africa 108.

Rhizoctonia bataticola (Taub.) Butl., 1925 = *Macrophomina phaseolina* (Tassi) Goid., 1950.

Rhizoctonia napae West.

India 518.

Rhizoctonia solani Kuehn = *Thanatephorus cucumeris* (Frank) Donk, 1956.

Rhizomorpha spp.

India 20.

West Pakistan 609.

Rhizopus sp.

Bulgaria 239.

Rhizopus nigricans Ehr.

Poland 612.

Sclerotinia p.

Bulgaria 239.

Sclerotinia libertiana Fckl. = *Whetzelinia sclerotiorum* (Lib.) Korf & Dumont, 1972.

Sclerotinia sclerotiorum (Lib.) de By., 1884 = *Whetzelinia sclerotiorum* (Lib.) Korf & Dumont, 1972.

Sclerotinia sclerotiorum (Lib.) Massee = *Whetzelinia sclerotiorum* (Lib.) Korf & Dumont, 1972.

Sclerotinia sclerotiorum (Lib.) Sacc. & Trott. = *Whetzelinia sclerotiorum* (Lib.) Korf & Dumont, 1972.

Sclerotium rolfsii Sacc., 1911.

India 73, 398.

Septoria sp.

Iowa 580.

Septoria argemones Tharp, 1917.

Nebraska 580.

Oklahoma 425, 426, 580.

Texas 560, 580.

Septoria chelidonii Desm., 1842.

Adzkarskaya ASSR 588.

Austria 211, 231, 564.

Belgium 469.

Bermuda 597.
Bulgaria 247, 298.
Caucasia, USSR 520.
China 340.
Czechoslovakia 409.
Denmark 279.
Finland 228.
France 212, 270, 466, 469, 585.
Germany 214.
Great Britain 382, 469.
Hungary 342.
Italy 469, 570.
Japan 503, 544, 545.
Karelian ASSR 277.
Latvian SSR 514.
Poland 341.
Portugal 542.
Puerto Rico 143, 537, 580.
Romania 485, 497.
Siberia 469.
Spain 287, 576, 578, 582.
Stavropol', USSR 361.
Switzerland 210.
Texas 580.
Tula Oblast, USSR 571.
Ukrainian SSR 275, 436.

Septoria rhoeadis Tassi, 1900.

Italy 477, 554.

Sphaerella arthopyrenioideis Auers., 1869 = *Mycosphaerella arthopyrenioideis* (Auers.) Larsen, 1932.

Sphaerella chelidonii Fautr. & Lambotte = *Mycosphaerella chelidonii* (Fautr. & Lambotte) Tomilin, 1969.

Sphaerella dendromeconis Cke. & Harkin., 1880.

California 467.
United States 126.

Sphaerella karajacensis Allesch., 1897 = *Mycosphaerella tassiana* (de N.) Johanson.

Sphaerella morphaea Sacc.

Italy 467.

Sphaerella schoenoprasi (Rab.) Auersw. as *Mycosphaerella* = *M. tassiana* (de N.) Johans., 1885.

Sphaerotheca macularis Magn. f. *papaveris* Simonian, 1958.

Armenian SSR 188, 521.

Sporotrichum pp.

India 20.

West Pakistan 609.

Stachylidium spp.

Italy 415.

Stemphylium alternariae (Cke.) Sacc.

Uzbek SSR and Tadzhik SSR borders 391.

Stemphylium botryosum Wallr. = *Pleospora herbarum* (Fr.) Rab., 1873.

Thanatephorus cucumeris (Fr.) Donk, 1956.

Syn.: *Pellicularia filamentosa* (Pat.) Rogers, 1943.

Rhizoctonia solani Kuehn.

Idaho 517, 580.

India 73, 581.

Indiana 580.

Kansas 580.

Maine 580.

New Jersey 580.

New York 580.

West Pakistan 609.

Trichoderma sp.

Poland 612.

Trichoderma viride Pers. ex Fr., 1829.

India 20.

West Pakistan 609.

Trichothecium sp. (sapro.)

Italy 415.

Trichothecium roseum Lk.

Bulgaria 239.

Germany 449.
Poland 612.

Verticillium spp.

Italy 155.

Verticillium albo-atrum Reinke & Berth.

California 580.
Germany 510, 605.
New York 580.

Verticillium dahliae Kleb.

New Zealand 55.

Whetzelinia sclerotiorum (Lib.) Korf & Dumont, 1972.

Syn.: *Sclerotinia libertiana* Fckl.

S. sclerotiorum (Lib.) de By., 1884.

S. sclerotiorum (Lib.) Sacc. & Trott.

S. sclerotiorum (Lib.) Massee

Bermuda 596, 597.
Crimea, USSR 305.
Germany 450.
Japan 138.
Kazakhstan, USSR 224.
Oregon 517.
Scotland 100, 134.
Sweden 450.
Tanganyika 453.

Yeast

Poland 612.

Bacteria

Bacillus aroideae Town., 1904 = *Erwinia aroideae* (Town.) Holland, 1920.

Bacillus papaveri Christoff [Khristov] in Atanasov, D. et al 1932 =
Erwinia aroideae (Town.) Holland, 1920.

Bacillus papaveris Ram Ayyar, 1927 = *Erwinia aroideae* (Town.) Holland, 1920.

Bacteria

Poland 612.

Bacterial soft rot

India 71.

Bacteriosis

Poland 141.

Bacterium carotovorum (Jones) Lehm. & Neuman, 1927.

Czechoslovakia 462.

Bacterium papavericola Bryan & McWhorter, 1930.

Bulgaria 234, 236.

Connecticut 65.

Japan 138.

Ohio 562.

Virginia 65.

Bacterium papaveris (Ram Ayyar) Burgv., 1935 = *Erwinia aroideae* (Town.)
Holland, 1920.

Bacterium papaveris Takimoto, 1935.

Japan 138, 549.

Bacterium papaverum ([Khristov] Christoff) Burgv., 1935.
Syn.: *Erwinia papaveris* Khristov 1933.

Bulgaria 21, 237.

Latvian SSR 514.

Yugoslavia 248, 332.

Erwinia aroideae (Town.) Holland, 1920.

Syn.: *Bacillus aroideae* Town., 1904.

B. papaveri Khristov in Atanasov, D. et al, 1932.

Bacillus papaveris Ram Ayyar, 1927.
B. papaveris (Ram Ayyar) Burgv., 1935.
B. papaveris (Ram Ayyar) Magrou in Hauduroy, 1937.

India 69, 78, 438.
Japan 138.
Taiwan 311.

Erwinia papaveris Christoff [Khristov], 1933 = *Bacterium papaverum*
(Khristov) Burgv., 1935.

Erwinia papaveris (Ram Ayyar) Magrou, 1937 = *E. aroideae* (Town.) Holland,
1920.

Pseudomonas spp.

Tanganyika 592, 593.

Pseudomonas papaveris Lelliott & Wallace, 1955.

India 523.
Tanganyika 278, 453.

Xanthomonas papavericola (Bryan & McWhorter) Dowson

Bulgaria 516.
Colombia 516.
Czechoslovakia 461, 462.
England 516, 528, 529.
Mauritius 111.
Romania 107, 516.
United States
 Arizona 580.
 Connecticut 84, 85, 580.
 Maryland 580.
 Massachusetts 47, 580.
 Missouri 580.
 New Jersey 580.
 New York 580.
 Ohio 580.
 Oregon 580.
 Texas 580.
 Virginia 580.

Nematodes

Acrobeloides buetschlii (de Man, 1884) Steiner & Buhner, 1933.

Kirghiz SSR 622.

Acrobeloides uberrinus Anderson, 1965.

Kirghiz SSR 622.

Aglenchus costatus (de Man, 1921) Andrassy, 1954.

Kirghiz SSR 622.

Alaimus primitivus de Man, 1880.

Kirghiz SSR 622.

Anguina sp.

Rhodesia & Nyasaland Fed. 301, 303.

Aphelenchoides sp.

Rhodesia & Nyasaland Fed. 301, 303.

Aphelenchoides fragariae (Ritzema Bos, 1891) Christie, 1932.

Kirghiz SSR 622.

Aphelenchoides parietinus (Bastian, 1865) Franklin, 1955.

Kirghiz SSR 622.

Aphelenchoides saprophilus Franklin, 1957.

Kirghiz SSR 622.

Aphelenchoides subparietinus Sanwal, 1961.

Kirghiz SSR 622.

Aphelenchoides subtenuis (Cobb, 1926) Steiner & Buhner, 1932.

Kirghiz SSR 622.

Aphelenchus avenae Bastian, 1865.

Kirghiz SSR 622.

Cephalobus persegnis Bastian, 1865.

Kirghiz SSR 622.

Chiloplacus symmetricus (Thorne, 1925) Thorne, 1937.

Kirghiz SSR 622.

Criconemoides tenuiannulata (Tulaganov, 1949) Zyubin, 1969.

Kirghiz SSR 622.

Diphtherophora communis (de Man, 1880) Goodey, 1963.

Kirghiz SSR 622.

Ditylenchus spp.

Rhodesia & Nyasaland Fed. 301, 303.

Ditylenchus destructor Thorne, 1945.

Kirghiz SSR 622.

Ditylenchus dipsaci (Kuehn, 1857) Filipjev, 1936.

Kirghiz SSR 622.

Dorylaimus filiiformis Bastian, 1865.

Kirghiz SSR 622.

Doryllium uniforme Cobb, 1920.

Kirghiz SSR 622.

Eucephalobus elongatus (de Man, 1880) Thorne, 1937.

Kirghiz SSR 622.

Eucephalobus oxyuroides (de Man, 1876) Steiner, 1936.

Kirghiz SSR 622.

Eucephalobus striatus (Bastian, 1865) Thorne, 1937.

Kirghiz SSR 622.

Eudorylaimus aculticauda (de Man, 1880) Andrassy, 1959.

Kirghiz SSR 622.

Eudorylaimus obtusicaudatus (Bastian, 1865) Andrassy, 1959.

Kirghiz SSR 622.

Eudorylaimus papillatus (Bastian, 1865) Andrassy, 1959.

Kirghiz SSR 622.

Eudorylaimus paraobtusicaudatus (Micol., 1922) Andrassy, 1959.

Kirghiz SSR 622.

Filenchus filiformis (Buetschli, 1873) Andrassy, 1954.

Kirghiz SSR 622.

Helicotylenchus multicinctus (Cobb, 1893) Golden, 1956.

Kirghiz SSR 622.

Helicotylenchus nannus Steiner, 1945.

Kirghiz SSR 622.

Heterodera marioni (Cornu) Goody

Israel 448.

Italy 74.

New South Wales 378.

United States 68.

Heterodera radicicola (Greef) Mueller

Germany 36.

Heterodera schachtii Schm., 1871.

Czechoslovakia 512.

Kirghiz SSR 622.

Hexatylus viviparus Goodey, 1926.

Kirghiz SSR 622.

Lelenchus infirmus Andrassy, 1954.

Kirghiz SSR 622.

Meloidogyne spp.

Israel 335.

Meloidogyne arenaria (Neal, 1889) Chitwood, 1949.

Rhodesia & Nyasaland Fec. 302.

Mesodiplogaster lheritieri (Maupas, 1919) Goodey, 1963.

Kirghiz SSR 622.

Mesodorylaimus bastiani (Buetschli, 1873) Andrassy, 1959.

Kirghiz SSR 622.

Monhystera agilis de Man, 1880.

Kirghiz SSR 622.

Monhystera bulbifera de Man, 1880.

Kirghiz SSR 622.

Mononchus papillatus (Bastian, 1865) Cobb, 1916.

Kirghiz SSR 622.

Neotylenchus abulbosus (Steiner, 1931) Goodey, 1933.

Kirghiz SSR 622.

Nothotylenchus acris Thorne, 1941.

Kirghiz SSR 622.

Panagrolaimus hygrophilus Bassen, 1940.

Kirghiz SSR 622.

Panagrolaimus rigidus (Schneider, 1866) Thorne, 1937.

Kirghiz SSR 622.

Panagrolaimus subelongatus (Cobb, 1914) Thorne, 1937.

Kirghiz SSR 622.

Paraphelenchus pseudoparietinus (Micol., 1922) Micol. 1925.

Kirghiz SSR 622.

Paraphelenchus tritici Baranovskaja, 1958.

Kirghiz SSR 622.

Plectus granulosus (Bastian, 1865) de Coninck & Stekh., 1933.

Kirghiz SSR 622.

Paratylenchus hamatus Thorne & Allen, 1950.

Kirghiz SSR 622.

Pratylenchus [undescribed sp.]

Netherlands 35.

Pratylenchus montanus Zubin, 1966.

Kirghiz SSR 622.

Pratylenchus penetrans (Cobb, 1917) Filipj. & Sch. Stkh., 1941.

Netherlands 35.

Pratylenchus scribneri Steiner, 1943.

Kirghiz SSR 622.

Pratylenchus vulnus Allen & Jensen, 1951.

Kirghiz SSR 622.

Prismatolaimus dolichurus de Man, 1880.

Kirghiz SSR 622.

Prismatolaimus intermedius (Buetschli, 1873) de Man, 1884.

Kirghiz SSR 622.

Rhabditis brevispina (Claus, 1862) Buetschli, 1873.

Kirghiz SSR 622.

Rhabditis intermedia de Man, 1880.

Kirghiz SSR 622.

Tetylenchus productus Thorne, 1949.

Kirghiz SSR 622.

Tylenchorhynchus dubius (Buetschli, 1873) Filipjev, 1936.

Kirghiz SSR 622.

Tylenchus sp.

Rhodesia & Nyasaland Fed. 301, 303.

Tylenchus agricola (de Man, 1884) Andrassy, 1954.

Kirghiz SSR 622.

Tylenchus davainei (Bastian, 1865) Andrassy, 1954.

Kirghiz SSR 622.

Viruses and Virus Suspects

Aster yellows

California 515.

Bean yellow mosaic virus

Bulgaria 263, 264.

Beet mosaic virus

Syn.: Poppy mosaic virus

Germany 179, 506, 507.

Poland 139.

Beet yellows virus

Netherlands 83, 291.

Big bud of tomato

New South Wales 185.

Black ring disease (no pathogen found)

Bangladesh [East Pakistan] 11.

Cabbage black ring spot virus

South Africa 109, 113, 322.

Curly top virus

Texas 12, 580.

Leaf crinkle disease

Bangladesh [East Pakistan] 11.

Spotted wilt

California 580.

New South Wales 201.

Tomato spotted wilt virus

California 384.

England 384, 527.
New South Wales 3, 4, 377.
New Zealand 54.
Queensland 184.
South Africa 109.
Tasmania 5, 6.

Virus [unidentified]

Czechoslovakia 460.
Ontario 292.

Yellows

New Brunswick, Canada 86.

Yellows virus

California 580.
Czechoslovakia 460.
Poland 139.
South Australia 484.

APPENDIX B--INDEX OF PATHOGENS AND THE HOSTS
UPON WHICH THEY ARE FOUND

Fungi

Aecidium plenum Arth.
Argemone mexicana L., 580^{6/}

Alternaria spp.
P. rhoeas L., 372.
P. somniferum L., 128, 334, 415, 448.

Alternaria alternata (Fr.) Keissler, 1912, 121 = *A. tenuis* Nees 1816/1817 =
Torula alternata Fr., 1832.
P. rhoeas L., 90.
P. somniferum L., 328.
P. spp., 452.

Alternaria brassicae (Berk.) Sacc., 1880 = *A. brunnea* = *A. somniferi* =
A. papaveris-somniferi = *Macrosporium bresadole* = *M. brassicae* = *M. brunnea*
= *M. commune* = *M. papaveris* = *M. somniferi*.
P. somniferum, 52, 53, 140, 141, 166, 205, 206, 207, 239, 242, 396, 397,
415, 474, 503, 504, 512, 526.

Alternaria lancipes Ellis & Ev., 1888.
Argemone mexicana L., 580.
Argemone platyceras Link & Otto, 120, 126, 474.

Alternaria tenuissima (Kunze ex Pers.) Wiltshire, 1933; syn. (Nees ex Fr.)
Wilts.
Argemone mexicana L., 552.

Aregma moniliiforme cf *Phragmidium*

Ascochyta chelidonii Kab. & Bub, 1907.
Chelidonium majus L., 480.

Ascochyta papaveris Oud., 1885 = *Diplodina papaveris* (Oud.) Lind, 280.
Ascigerous stage is *Didymella*.
P. nudicaule L., 109, 474.
P. radicum Rottb., 280, 281.

Ascochyta sp.
P. somniferum, 62.

^{6/} Numbers after hosts refer to entries in the bibliography.

Aspergillus sp. *Micheli* ex Fries (saprophyte).

P. somniferum L., 449.

Aureobasidium pullulans (de By.) Arnaud. = *Pullularia pullulans*, 121.

P. somniferum L., 612.

Botryotinia (*Sclerotinia*) *fuckeliana* (de By.) Whet. Imperfect state is

Botrytis cinerea Pers. ex Fr.

Eschscholtzia californica Cham., 580.

P. nudicaule L., 54, 580.

P. orientale L., 580.

P. radicans Rottb., 280.

P. rhoeas L., 580.

P. somniferum L., 138, 354, 429, 450.

P. spp., 13, 141.

Botrytis sp.

P. somniferum L., 450.

Botrytis arborescens cf *Peronospora arborescens*.

Botrytis cinerea cf *Botryotinia*.

Brachycladium penicillatum Corda cf *Pleospora papaveracea*.

Caeoma chelidonii Magn.

Chelidonium majus L., 421, 471.

Cercospora sp.

P. sp., 531.

Cercospora papavericola Chupp, 1954 comb. nov. = *C. papaveri* Mueller & Chupp, 1936; *C. papaveri* Nakata; *C. papaveris* Saw. nom. nud.

Argemone mexicana L., 233.

P. pavoninum Fisch. & C. A. Mey, 225.

P. rhoeas L., 401.

P. somniferum L., 81, 138, 402, 504, 606, 609.

P. sp., 356, 538.

Cercospora whetzelii Chupp in Toro, 1931.

Argemone mexicana L., 81, 569, 580.

Argemone platyceras Link & Otto, 81.

Cercosporidium guanicensis (Stev.) Deighton, 1967 = *Cladosporium guanicensis* (Stev.) Deighton, 1917 (sapro.).

Argemone mexicana L., 531, 537, 539, 580.

Chaetomella atra Fckl.

P. rhoeas L., 10, 392.

- Cladosporium* sp.
P. somniferum, 128, 137.
- Cladosporium cladosporioides* (Fresen.) de Vries, 1952.
Argemone mexicana L., 552.
- Cladosporium herbarum* cf. *Mycosphaerella tassiana*.
- Clathrospora pentamera* (Karst.) Berl., 1900 nom. cons. prop. vs *Pleospora*.
P. radicum Rottb., 280, 281.
- Colletotrichum circinans* (Berk.) Arx.
Eschscholtzia californica Cham., 217.
- Cucurbitaria papaveracea* cf. *Pleospora papaveracea*.
- Curvularia geniculata* (Tracy & Earle) Boedijn, 1933 (*Cochliobolus geniculata* (Nelson)).
P. somniferum L., 154.
- Dacrymyces* (*Dacryomyces*) *papaveris* Quel., 1892.
P. somniferum L., 430, 475.
- Dactylium roseum* Berk. (sapro) = *Tricothecium roseum* Link, 20.
P. somniferum L., 20, 609.
- Dematium pullulans* de By.
P. sp., 321.
- Dendryphon* (*Dendryphium*) *penicillatum* cf. *Pleospora papaveracea*.
- Dendryphium papaveris* cf. *Pleospora papaveracea*.
- Dendryphium ramosum* cf. *Pleospora papaveracea*.
- Didymaria chelidonii* Jacz. in Komarov, 1900.
Chelidonium uniflorum Sieb. & Zucc., 254, 477.
- Diplodina eschscholtziae* Oudem., 1904.
Eschscholtzia crocea, 479.
- Diplodina papaveris* cf. *Ascochyta papaveris*.
- Diplodirula glaucii* (Cke. & Mass.) Fl. Tassi, 1902.
Glaucium flavum Crantz., 555.
- Drechslera tetramera*.
P. somniferum L., 79.

Entyloma sp.

P. dubium L., 62.

Entyloma bicolor Zopf. in Rab., 1878 cf. *E. fuscum*.

Entyloma chelidonii Cif.

Chelidonium majus L., 285.

Entyloma eschscholtziae Harkn.

Eschscholtzia californica Cham., 126.

Entyloma fuscillum Rab. cf. *E. fuscum*.

Entyloma fuscum Schroet. = *E. bicolor* Zopf. in Rab.; *E. bicolor* Rab., 1878
nom. nud., 82; *E. fuscillum*.

Glaucium flavum Crantz., 488.

P. argemone L., 82, 285, 344, 471.

P. atlanticum (Ball.) Coss., 488.

P. commutatum Fisch. & C. A. Meyer, 49, 153.

P. dubium, 44, 82, 279, 284, 285, 288, 344.

P. glaucum Boiss & Hausskn., 488.

P. litvinovii Fedde & Bornm., 223, 256.

P. monanthum Trautv., 488.

P. nudicaule L., 276, 285.

P. orientale L., 488.

P. pavoninum Fisch. & C. A. Meyer, 225.

P. rhoeas L., 8, 9, 18, 39, 44, 82, 109, 147, 250, 284, 295, 315, 316,
317, 320, 344, 359, 363, 375, 386, 392, 433, 471, 483, 488, 575, 577,
586, 596.

P. setigerum D. C., 488.

P. somniferum, 44, 49, 50, 56, 60, 64, 82, 107, 128, 141, 145, 153, 226,
242, 250, 284, 300, 326, 357, 363, 412, 488, 596.

P. spp., 73, 314, 437, 452, 490.

Epicoccum herbarum Cda. (sapro)

P. somniferum.

Erysiphe spp.

P. rhoeas L., 274.

P. somniferum L., 226, 379, 519.

Erysiphe cichoracearum CC ex Merat. = *E. communis* Grev. = *E. communis*

(Wallr.) Blum. = *E. martii* Lev. Ref. Blumer 1967:230 = *E. cichoracearum*

DC em. Salm. f. *papaveris* Pot.

Chelidonium majus L. var. *asiaticum*, 188.

Eschscholtzia californica Cham., 188.

Meconopsis cambrica Vig., 188.

P. alpinum L., 188.

P. amurense Hort. ex Karrer, 188.

P. angrenicum Panzij, 391.

- P. argemone* L., 63, 188.
- P. dubium* L., 188, 320.
- P. dubium* var. *lecoquii* Lamotte, 188.
- P. hybridum* L., 188.
- P. nudicaule* L., 188, 319, 320.
- P. rhoeas* L., 188, 320.
- P. somniferum* L., 44, 63, 73, 94, 95, 107, 138, 157, 188, 224, 225, 320, 398, 422, 423, 443, 499, 500, 511.

Erysiphe polygoni DC

- Chelidonium majus* var. *asiaticum* L., 189.
- Eschscholtzia californica* Cham., 186.
- P. dubium* L., 37, 440, 441.
- P. nudicaule* L., 7, 432.
- P. rhoeas* L., 316.
- P. somniferum* L., 22, 64, 72, 73, 78, 80, 128, 176, 177, 186, 242, 257, 262, 315, 517.

Fusarium spp.

- P. somniferum* L., 141, 239, 448.

Fusarium concolor Reinking

- P. somniferum* L., 612.

Fusarium equiseti (Cda.) Sacc.

- P. rhoeas* L., 159.

Fusarium martii Appel & Wor.

- P. somniferum* L., 367.

Fusarium oxysporum Schl. emend. Snyder & Hans.

- P. rhoeas* L., 159.

Fusarium oxysporum Schlecht. f. *cubense* E. F. Smith Wr.

- P. somniferum* L., 367.

Fusarium scirpi Lamb. & Fautr., 1894.

- P. nudicaule* L., 108, 109.
- P. rhoeas* L., 108, 109.

Fusarium scirpi Lamb. & Fautr. f. *caudatum* Wr.

- P. somniferum* L., 128, 239.

Fusarium scirpi f. *compactum*.

- P. rhoeas* L., 108.

Fusarium solani (Maet.) Appel & Wr. em Snyder & Hans.

- P. rhoeas* L., 159.

- Glomerella cingulata* (Ston.) Spauld. & Schrenk. Imperfect state =
Colletotrichum gloeosporioides (Penzig.) Sacc. (v. Arx lists 600 synonyms)
 Ref.: J. A. von Arx, 1970. = *Gloeosporium argemones* Ellis and Everh, 1887.
Argemone platyceras Link & Otto, 119, 126, 474, 541.
P. orientale L., 405.
P. somniferum L., 180, 181, 182, 612.
- Helicobasidium purpureum* (Tul.) Pat. Imperfect state: *Rhizoctonia crocorum*
 DC ex Fr.
P. orientale L., 406.
- Heteropatella lacera* (Fckl.)
P. alpinum L., 360.
- Heteropatella umbilicata* (Fr.) Jaap.
P. radicum Rottb., 281.
- Heterosporium echinulatum* cf. *Mycosphaerella dianthi*.
- Heterosporium eschscholtziae* Harkn.
Eschscholtzia californica Cham., 126, 470, 580.
- Heterosporium groenlandicum* Allesch.
P. nudicaule L., 109, 476.
P. somniferum L., 423.
- Hypocrea spinulosa* Fckl.
Chelidonium majus L., 468.
- Leptosphaeria chelidonii* Fautr., 1895.
Chelidonium majus L., 476.
- Leptosphaeria papaveris* Rostr., 1903.
P. radicum Rottb., 271, 478.
- Leptosphaeria pellita* (Rab. & Klotzsch.) Sacc.
P. somniferum L., 466.
- Leveillula papaveracearum* Golov. f. *argemones* Golov f. nov., 1956.
Argemone platyceras Link & Otto, 156, 188.
- Leveillula taurica* (Lev.) Arn. = *Erysiphe taurica* Lev.
Argemone mexicana L., 188, 233, 552.
Argemone platyceras Link & Otto, 188.
Eschscholtzia sp., 187, 188, 453.
P. rhoeas L., 188, 233.
P. somniferum L., 188.
P. spp., 187.

- Lophodermium svalbardense* Lind., 1928.
P. radicatum Rottb., 280.
- Macrosporium* spp. cf. *Alternaria*.
- Macrophoma papaveris* Camara, 1930.
P. somniferum L., 75.
- Macrophomina phaseoli* (Maubl.) Ashby, 1927 = *M. phaseolina* (Tassi.) Goid.
 1950 = *Rhizoctonia bataticola* (Taub.) Butl.
P. somniferum L., 78, 101.
- Massaria cleistotheca* Harkn.
Dendromecon rigidum Benth., 126, 473.
- Melampsora chelidoni-pieroti* Matsu., 1926.
Corydalis incissa Pers., 310.
- Melampsora larici-populina* Kleb.
Chelidonium sp., 514.
- Melampsora magnusiana* Wagner.
Chelidonium majus L., 190, 208, 246, 279, 309, 310, 333, 388, 446, 520,
 547.
- Metasphaeria patelliformis* Kirsch., 1935.
Chelidonium majus L., 244.
- Mucilago spongiosa* (Leyss.) Morg. (Myxomycete, saprophyte).
Meconopsis spp., 100, 134.
- Mucor mucedo* (L.) Lk. (saprophyte)
Mucor mucedo Fr.
P. somniferum L., 20, 239.
- Mycosphaerella chelidonii* (Fautr. & Lambotte) Tomilin, 1969. *Sphaerella chelidonii* Sydowia 3: 41.
Chelidonium majus L., 476.
- Mycosphaerella dianthi* (Burt.) Jørstad, 1945. Imperfect state is
Cladosporium echinulatum (Berk.) de Vries, 1952. *Heterosporium echinulatum* (Berk.) Cooke, 1877.
P. somniferum L., 239.
- Mycosphaerella pachyasa* (Fckl.) Berl. & Vogl.
P. nudicaule L., 98.

Mycosphaerella tassiana (de N.) Johans., 1885.

Mycosphaerella arthopyrenioides (Auers.) Larsen, 1932.

Physalospora polaris Rostr.

Sphaerella arthopyrenioides Auers., 1869, Sydowia 3: 40.

Sphaerella schoenoprasii

Imperfect state is *Cladosporium herbarum* (Pers.) Link S. F. Gray, 1821.

P. burseri Crantz, 467.

P. croceum Ledeb., 223, 256.

P. litvinovii Fedde & Bornm., 223, 256.

P. nudicaule L., 126, 411, 413, 473.

P. radicum Rottb., 271, 276, 280, 281.

P. somniferum L., 20, 135, 141, 225, 268, 277, 328, 422, 428, 449, 489, 609.

Mycosphaerella tulasnei Jancz.

P. somniferum L., 239.

Myrothecium Tode ex Fr. sp.

Argemone mexicana L., 597.

Oidiopsis sp.

P. somniferum L., 258.

Oidiopsis papaveris Saw., 1933.

P. somniferum L., 501.

Oidium spp.

Argemone mexicana L., 188, 398.

Chelidonium majus var. *grandiflorum* L., 188.

C. luteum Gilib., 188.

P. nudicaule L., 54, 188, 197.

P. orientale L., 188.

P. rhoeas L., 63, 188, 453.

P. somniferum L., 19, 188.

P. spp., 188, 195, 196.

Oidium erysiphoides Fr. f. *papaveris* Roum.

Chelidonium majus L., 482, 502.

P. humifusum Fedde., 456.

P. orientale L., 238.

P. rhoeas L., 73, 338, 399.

P. somniferum L., 238, 268, 423, 530.

Ophiobolus sativus (P.K.B.) Ito & Kurib.

P. somniferum L., 239.

Ophiobolus tenellus (Awd.) Sacc.

Chelidonium majus L., 279.

P. rhoeas L., 91.

Ovularia Sacc. sp.

Argemone platyceras, 531.

Ovularia indica R. Rao, 1968. Ascigerous stage is *Mycosphaerella*.

Argemone mexicana L., 442.

Pellicularia filamentosa (Pat.) Rogers.

P. somniferum, 73.

Penicillium spp. (saprophyte).

P. somniferum, 238, 612.

Peronospora arborescens = *Botrytis arborescens* = *Peronospora effusa* f. *papaveris* = *Peronospora grisea* = *Peronospora papaveris*.

Argemone mexicana L., 71, 72, 145, 308, 338, 399, 424, 543, 580.

Argemone platyceras Link & Otto, 200, 603 (incorrectly reported as *P. corydalis*)

Chelidonium majus L., 199.

Meconopsis betonicifolia Franch., 67, 89.

Meconopsis cambrica Vig., 89, 145, 315.

Meconopsis grandis Prain., 89.

Meconopsis integrifolia Franch., 89.

Meconopsis latifolia (Prain.) Prain., 89.

Meconopsis paniculata (D. Don.) Prain., 89.

Meconopsis prattii Prain., 89.

Meconopsis superba King ex Prain., 89.

Meconopsis napaulensis DC (*M. wallichii* Hook.), 89.

M. spp., 100, 134, 370, 406.

P. alpinum, 145, 339, 387, 406, 548.

P. argemone, 34, 66, 71, 89, 127, 145, 198, 279, 293, 345, 466, 584.

P. canadense, 67.

P. commutatum, 48, 153.

P. dubium, 34, 66, 67, 89, 100, 127, 134, 145, 169, 171, 199, 209, 213, 215, 272, 273, 279, 293, 297, 320, 345, 385, 386, 389, 407, 439, 452, 471, 492, 494, 525, 584.

P. floribundum Desf., 67.

P. hybridum L., 296, 297, 452.

P. nudicaule L., 3, 4, 67, 283, 377, 563.

P. orientale L., 320.

P. "pannoicum", 67.

P. polare (Tolm.) Perfiljco, 67.

P. pavoninum Fisch. & C. A. Mey, 223, 225, 256, 391.

P. rhoeas L., 8, 34, 44, 67, 71, 99, 127, 136, 145, 169, 171, 199, 279, 293, 295, 297, 314, 316, 317, 320, 343, 345, 390, 392, 400, 416, 431, 439, 444, 452, 454, 457, 471, 481, 492, 524, 525, 553, 561.

P. setigerum, 26, 297.

- P. somniferum*, 1, 2, 8, 19, 20, 25, 26, 27, 32, 33, 34, 40, 42, 43, 46, 48, 57, 66, 67, 70, 71, 72, 73, 78, 87, 89, 92, 94, 95, 99, 100, 103, 106, 107, 114, 116, 127, 128, 134, 135, 137, 138, 139, 141, 145, 151, 167, 169, 170, 172, 174, 178, 183, 199, 202, 203, 204, 206, 215, 219, 220, 221, 222, 225, 226, 227, 239, 242, 243, 248, 249, 253, 257, 259, 261, 265, 279, 282, 286, 287, 289, 293, 306, 307, 308, 312, 315, 320, 327, 332, 339, 343, 346, 347, 354, 361, 365, 366, 379, 380, 383, 386, 387, 390, 391, 394, 395, 399, 410, 414, 417, 418, 422, 427, 441, 452, 471, 481, 492, 495, 505, 508, 512, 513, 525, 532, 533, 534, 543, 548, 565, 567, 573, 574, 579, 609, 611, 621.
- P. spp.*, 41, 77, 87, 92, 158, 162, 216, 294, 306, 314, 393, 417, 490, 514, 540, 546, 583, 588.

Peronospora arborescens, ultraviolet light, influence on conidia of.

Peronospora argemone Gaeum.

Chelidonium majus L., 376.

Eschscholtzia californica Cham., 376.

P. argemone L., 169, 171, 390, 444, 525.

P. croceum Ledeb., 225.

P. dubium, 272, 273, 279.

P. rhoeas L., 376.

P. somniferum L., 455.

Peronospora chelidonii Miy.

Chelidonium majus L., 191, 387, 545, 566.

Peronospora cristatum Tranz.

P. hybridum L., 390, 444.

Peronospora gaeumanni Mayor, 1949 = *P. meconopsidis* Mayor, 1958 nom. nov.

P. gaeumanni Mund., 1938.

Argemone mexicana L., 148, 233, 358, 399, 403.

Meconopsis cambrica Vig., 318.

Peronospora glaucii Savul. & Rayss, 1935 = *P. glaucii* Lob.

Glaucium corniculatum, 492, 496.

Peronospora meconopsidis

Meconopsis cambrica Vig., 320.

Peronospora parasitica de By.

P. sp., 393.

Peziza eschscholtziae Phill & Harkn.

Eschscholtzia californica Cham., 126.

Phialea cyathoides (Bull.) Gill.

Chelidonium majus L., 472.

- Phialea eschscholtziae* (Phill. & Harkn.) Sacc., 1889.
Eschscholtzia californica Cham., 472.
- Phoma melaena* (Fr.) Mont. & Dur.
P. somniferum L., 51.
- Phoma morphea* Sacc., 1881.
P. somniferum L., 465, 469.
- Phoma rhoeadis* Brun.
P. alpinum L., 369.
P. dubium L., 279.
P. glaucum Boiss. & Hausskn., 369.
P. nudicaule L., 90, 369.
P. orientale L., 369.
P. paeoniflorum Hort. ex Correa, 369.
P. rhoeas L., 475.
P. umbrosum Hort., 90.
- Phoma sphaeronaemoides* Fautr., 1895.
Chelidonium majus L., 476.
- Phoma striaeformis* Dur. & Mont. f. *hysteriola* Sacc.
P. somniferum L., 61, 466, 469.
- Phragmidium mucronatum* (Pers.) Schlecht. = *Aregma moniliiforme*.
P. somniferum L., 20.
- Phyllosticta chelidonii* Bres., 1896.
Chelidonium majus L., 476.
- Phymatotrichum omnivorum* (Shear.) Duggar, 1916.
Argemone mexicana L., 580.
Argemone sp., 240, 613.
Chelidonium majus L., 580.
- Physalospora polaris* cf. *Mycosphaerella tassiana*.
- Phytophthora cactorum* (Leb. & Cohn) Schroet., 1886.
Chelidonium majus L., 376.
Eschscholtzia californica Cham., 376, 572.
Meconopsis sp., 134.
Meconopsis napaulensis DC. (*M. wallichii*), 100.
P. argemone L., 376.
P. rhoeas L., 376.
- Phytophthora cryptogea* Pethybr. & Lasf., 1919.
P. nudicaule L., 3, 6, 59, 378, 572.

Phytophthora nicotiana f. *parasitica* (Dastur) comb. nov., 1963.

Eschscholtzia californica Cham., 602.

Meconopsis spp., 100, 134.

Phytophthora verrucosa Alcock & Foister in Foister, 1940.

Meconopsis spp., 100, 133, 134, 595.

Phytophthora spp.

P. nudicaule L., 58, 76, 572.

Plasmodiophora brassicae Wor.

R. rhoeas L., 252.

Pleosphaerulina californica Berl.

Dendromecon rigidum Benth., 476, 580.

Pleospora argyrospora Harkn.

Dendromecon rigidum Benth., 126, 473.

Pleospora herbarum (Pers. ex Fr.) Rab. Imperfect state is *Stemphylium botryosum* Wallr., 1833 (Ellis, 121).

Chelidonium majus L., 279.

P. argemone L., 445.

P. monanthum, 587.

P. nudicaule L., 98, 126, 276.

P. radicum Rottb., 280.

P. somniferum L., 354.

P. spp., 279.

Pleospora media

P. nudicaule L., 580.

Pleospora papaveracea = *P. calvescens* = *Pyrenophora calvescens* = *Sphaeria calvescens* = *sphaeria pellita* = *Cucurbitaria papaveracea*.

Imperfect state is *Dendryphion penicillatum* = *Dendryphium penicillatum* =

Helminthosporium papaveris = *H. papaveri* = *Dendryphium papaveris* =

D. ramosum = *D. penicillatum* f. *sclerotiale*.

Chelidonium majus L., 470.

P. alpinum L., 90, 369.

P. glaucum Boiss. & Hausskn., 373.

P. mursellii Hort., 90, 368.

P. nudicaule L., 90, 126, 369, 371.

P. orientale L., 522.

P. paeoniflorum Hort. ex Correa, 90, 368, 373.

P. rhoeas, 90, 91, 245, 331, 368, 371, 373, 374, 439, 458, 466, 468, 470.

P. somniferum, 19, 23, 24, 25, 30, 31, 38, 45, 90, 91, 93, 94, 95, 96, 97,

102, 107, 112, 115, 121, 123, 128, 129, 130, 131, 132, 139, 144, 150,

152, 154, 163, 164, 165, 167, 168, 178, 206, 226, 230, 234, 235, 239,

242, 245, 248, 251, 289, 290, 304, 307, 313, 324, 325, 330, 332, 336,

337, 348, 349, 350, 351, 352, 353, 354, 355, 357, 368, 373, 374, 379, 380, 395, 404, 408, 419, 420, 422, 434, 435, 447, 448, 449, 450, 451, 452, 468, 486, 498, 504, 509, 510, 512, 533, 534, 550, 598, 604, 610, 612, 616, 617, 618.

P. umbrosum Hort., 90, 369.

P. spp., 90, 149, 198, 370, 381, 385, 386, 459, 468, 514.

Pleospora papaveracea, antagonistic microorganisms to, 97.

Pleospora papaveracea, antibiotics, effect of on, 110, 124, 131, 559.

Pleospora papaveracea, actinomycetes, reaction to, 364.

Dendryphion penicillatum, disinfectants, effect on conidia of, 349.

Dendryphion penicillatum, fungicides, effect on conidia of, 129, 132.

Pleospora scrophulariae (Desm.) Hoehn.

P. radicum Rottb., 281.

Pleospora vulgaris Niessl = *P. infectoria* Fckl.

P. nudicaule L., 98.

P. rhoeas L., 91.

Pleosphaerulina californica

Dendromecon rigidum Benth., 476, 580.

Dendromecon sp., 480.

Pocosphaeria dendromeconis Earle, 1904.

Dendromecon rigidum Benth., 580.

Pullularia pullulans cf. *Aureobasidium pullulans* (de By.) Arnaud

P. somniferum L., 612.

Pyrenophora androsaces (Fckl.) Sacc.

P. radicum Rottb., 280.

Pyrenophora chrysospora (Niessl.) Sacc. f. *polaris* Karst., 1884 = *P. cerastii* (Oud.) Lind, 1924.

P. nudicaule L., 473.

P. radicum Rottb., 280, 281.

Pyrenophora paucitricha (Fckl.) Berl. & Vogl.

P. nudicaule L., 98.

Pythium spp.

P. nudicaule L., 108.

P. somniferum L., 17.

- Pythium deBaryanum* Hesse, 1874.
Eschscholtzia californica Cham., 329.
- Pythium mamillatum* Meurs, 1928.
P. somniferum L., 15, 16.
- Pythium megalacanthum* de By., 1881.
P. nudicaule L., 329.
P. rhoeas L., 558.
- Pythium oligandrum* Drechs., 1930.
P. rhoeas L., 109, 329, 590.
- Pythium spinosum* Saw., 1926.
P. rhoeas L., 329.
- Pythium ultimum* Trow., 1901.
P. nudicaule L., 109, 269, 329, 568, 590.
P. somniferum L., 14, 15, 16.
- Rhabdospora rhoeadis* Tassi, 1904.
P. rhoeas L., 479, 556, 557.
- Rhizoctonia napi*.
Argemone mexicana L., 518.
- Rhizoctonia solani* cf. *Thanatephorus cucumeris*.
- Rhizomorpha* sp.
P. somniferum L., 20, 609.
- Rhizopus nigricans* Ehr. = *R. stolonifer* (Ehr. ex Fr.) Lind (saprophyte).
P. somniferum L., 612.
- Rhizopus* sp. (saprophyte).
P. somniferum L., 239.
- Sclerotium rolfsii* Sacc.
P. somniferum, 73, 398.
- Sclerotinia* cf. *Whetzelinia*.
- Septoria argemone* Tharp, 1917.
Argemone alba Lestib., 426.
Argemone intermedia Sweet, 426.
Argemone mexicana L., 425, 580.
Argemone platyceras Link & Otto, 560.

Septoria chelidonii Desm., 1842.

Argemone mexicana L., 143, 537, 539, 580, 597.

Chelidonium majus L., 210, 211, 212, 214, 228, 229, 231, 247, 270, 275,
277, 287, 298, 340, 341, 342, 361, 382, 409, 423, 436, 466, 469, 485,
497, 503, 520, 542, 544, 545, 564, 570, 571, 576, 578, 580, 582, 584,
585.

Septoria rhoeadis Tassi, 1900.

P. rhoeas L., 477, 554.

Septoria spp.

P. sp., 580.

Sphaerella dendromeconis.

Dendromecon rigidum Benth., 126, 467.

Sphaerella karajacensis.

P. nudicaule L., 476.

Sphaerella morphaea Sacc.

P. somniferum L., 467.

Sphaerotheca macularis Magn. f. *papaveris* Simonian, 1958.

P. orientale L., 188, 521.

Sporotrichum Link ex Fr., 1832 sp.

P. somniferum L., 20, 609.

Stachylidium Link ex S. F. Gray; Link, 1809 sp. (saprophyte).

P. somniferum L., 415.

Stemphylium alternariae (Cks.) Sacc.

P. pavoninum Fisch. & C. A. Mey, 391.

Stemphylium botryosum cf. *Pleospora herbarum*.

Thanatephorus cucumeris (Frank) Donk.

Rhizoctonia solani Kuehn. Ref.: von Arx, 1970.

P. nudicaule L., 580.

P. orientale L., 580.

P. rhoeas L., 580.

P. somniferum L., 580.

Thanatephorus spp.

P. nudicaule L., 108, 109.

P. orientale L., 142.

P. rhoeas L., 108, 109.

P. somniferum L., 70, 71.

Thielaviopsis basicola (Berk. & Br.) Ferraris, 1912.

P. nudicaule L., 218.

Torula herbarum (Pers.) Link ex S. F. Gray, 1821.

P. nudicaule L., 470.

Trichoderma viride Pers. ex Fr., 1829.

P. somniferum L., 20, 609.

Trichoderma sp.

P. somniferum L., 612.

Trichothecium sp. (sapro.)

P. somniferum L., 415.

Trichothecium roseum Link ex Fr. = *Dactylium roseum*.

P. somniferum L., 239, 449, 612.

Verticillium sp.

P. orientale L., 104, 323.

P. pilosum Sibth. & Sm., 105.

P. rhoeas L., 104, 155.

P. somniferum L., 128, 355.

Verticillium albo-atrum Reinke & Berth.

Eschscholtzia californica Cham., 463, 580.

P. bracteatum Lindl., 605.

P. orientale L., 463, 580.

P. rhoeas L., 463.

P. somniferum L., 510.

Verticillium dahliae Kleb.

Eschscholtzia californica Cham., 323.

P. nudicaule L., 55.

P. orientale L., 323.

P. rhoeas L., 323.

Whetzelinia sclerotiorum Korf and Dumont = *Sclerotinia sclerotiorum* (Lib.)

de By. = *Sclerotinia libertiana* Fckl.

Argemone mexicana L., 596, 597.

Eschscholtzia sp., 453, 594.

Meconopsis spp., 100, 134.

P. somniferum L., 138, 224, 305, 354, 450, 517.

Bacteria

Erwinia aroidea = *Bacillus aroidea* = *Bacillus croci* = *Bacillus papaveris* =
Bacterium aroidea = *Bacterium papaveris* = *Erwinia croci* = *Erwinia melonis* =
Erwinia papaveris = *Pectobacterium aroidea* = *Pectobacterium melonis*.

Argemone mexicana L., 175, 438.

P. alpinum L., 21, 117, 237.

P. orientale L., 21, 117, 141, 237, 549.

P. rhoeas L., 117, 175, 438, 549.

P. somniferum L., 21, 69, 78, 117, 128, 138, 141, 226, 237, 248, 311,
332, 355, 461, 535, 549.

P. sp., 514, 535.

production of pectic enzymes by, 528, 529.

Erwinia carotovora (Jones, 1901) Holland, 1920 = *Bacillus carotovorus* =
Bacterium carotovorus.

P. somniferum L., 71, 354.

P. sp., 462.

Phytomonas tabaca Wolff & Foster = *Pseudomonas tabaci* (Wolf & Foster)
Stevens .

P. somniferum L., 28.

Pseudomonas papaveris Lelliott & Wallace, 1955.

P. orientale L., 278, 523.

P. rhoeas L., 278, 453, 523.

Pseudomonas spp.

P. rhoeas L., 592, 593.

Xanthomonas papavericola (Bryan & McWhorter, 1930) Dowson, 1939 =
Bacterium papavericola = *Phytomonas papavericola*.

Argemone sp., 111, 516.

Eschscholtzia californica Cham., 580.

Meconopsis baileyi, 516.

P. orientale L., 47, 65, 85, 111, 516, 535, 580, 601.

P. rhoeas L., 65, 84, 85, 516, 535, 580, 601.

P. somniferum L., 107, 138, 234, 236, 299, 354, 461, 516, 580.

P. spp., 85, 111, 160, 462, 562, 580.

nutritive requirements of, 536.

production of pectic enzymes by, 528, 529.

serological studies on, 122.

Viruses

Anemone Mosaic

P. rhoeas L., 192.

P. somniferum L., 192.

Aster Yellows (a mycoplasma)

Eschscholtzia californica Cham., 266, 515.

P. nudicaule L., 267.

Bean Yellow Mosaic Virus

P. somniferum, 263, 264.

Beet Curly Top Virus

P. nudicaule L., 12, 580.

P. orientale L., 12, 580.

Beet Mosaic Virus

Argemone intermedia Sweet, 506.

Argemone mexicana L., 506.

Argemone platyceras Link & Otto, 506.

P. rhoeas L., 506.

Big Bud of Tomato

Argemone mexicana L., 185.

Cabbage Black Ring Spot Virus

P. nudicaule L., 109, 113, 173, 322.

P. rhoeas L., 109, 113, 173, 322.

P. somniferum L., 11, 109, 113, 173, 322.

Callistephus Virus 1 and 1A

Eschscholtzia californica Cham., 580.

Cucumber Mosaic Virus

Argemone mexicana L., 255.

P. orientale L., 255.

P. rhoeas L., 255.

Datura 437 Virus

P. somniferum L., 93A.

Leaf Crinkle

P. somniferum L., 11.

Pelargonium Leaf Curl

P. somniferum L., 193.

Poppy Mosaic

P. somniferum L., 139, 179, 354, 507.

Sugar Beet Yellow

Eschscholtzia californica Cham., 29.

P. nudicaule L., 484.

P. rhoeas L., 614.

P. somniferum L., 29, 83, 291, 464.

P. sp., 29, 86.

Tomato Spotted Wilt

P. nudicaule L., 3, 4, 5, 6, 54, 109, 184, 201, 377, 384, 527, 580.

Virus?

Chelidonium sp., 292.

P. somniferum L., 232.

Yellowing Virus

P. somniferum L., 139, 354, 460.

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